



Assessment of clinical pathology, pathogen exposure, and impact of mercury in two reintroduced populations of American marten in Michigan

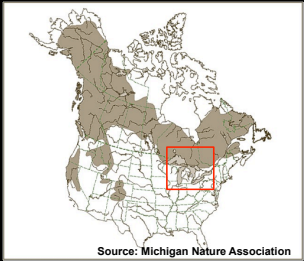
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Room 160 PBB, 12:20 P.M., March 20, 2013

Introduction: *Martes americana*

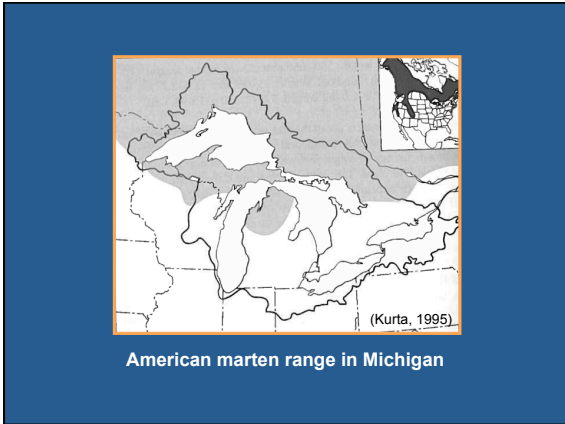
- Family Mustelidae
- Males 900-1300 gm
- Females 600-900 gm
- Mesocarnivore
- Forest habitat
- 1-4 kits born in spring

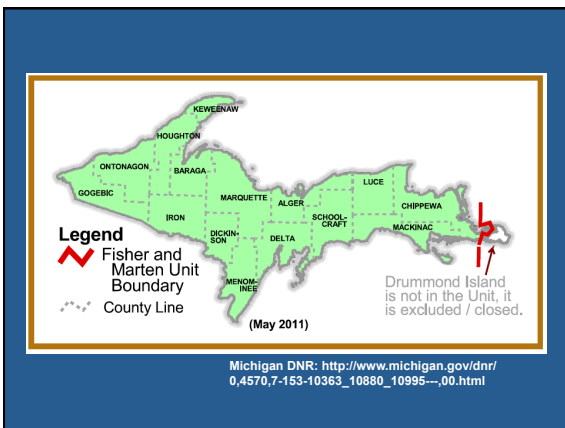


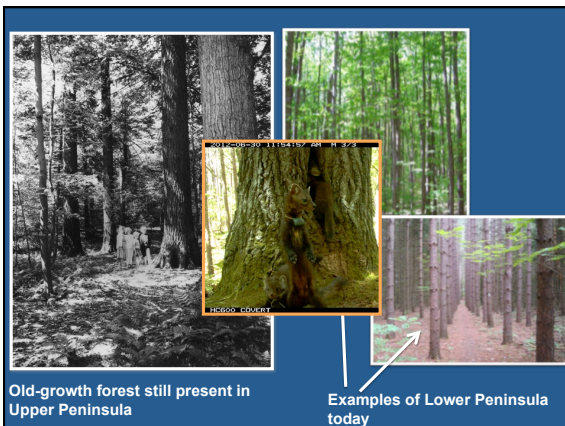
North American distribution of marten



Source: Michigan Nature Association







Collaborative Research



Disease in marten in Michigan?

Veine-Smith, A.M., J. Bird, and J.L. Belant. 2011. Patterns of endoparasite infections in American martens (*Martes americana*) of the Upper Peninsula of Michigan, U.S.A. *Comparative Parasitology* 78: 225-232.



Expanding the search:

Bourque, M. 1985. A survey of *Trichinella spiralis* in wild carnivores in Southwestern Quebec. *The Canadian Veterinary Journal* 26: 203-204.

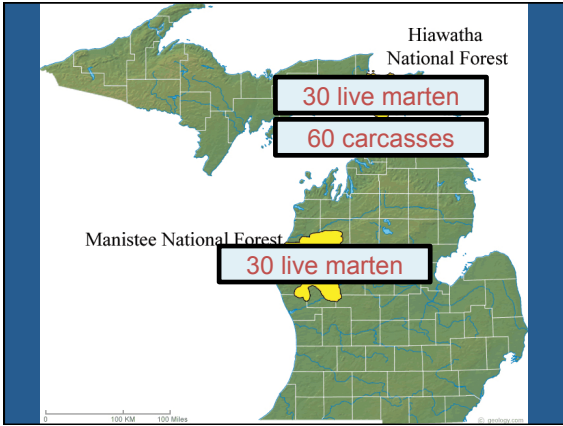
Bremle, G., P. Larsson, and J.O. Hellin. 1997. Polychlorinated biphenyls in a terrestrial predator, the pine marten (*Martes martes*). *Environmental Toxicology and Chemistry* 16: 1779-1784.

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Frollich, K., O. Czupalla, L. Haas, J. Hentschke, J. Dedek, and J. Fickel. 2000. Epizootiological investigations of canine distemper virus in free-ranging carnivores from Germany. *Veterinary Microbiology* 74: 283-292.

Frollich, K., W.J. Streich, J. Fickel, S. Jung, U. Truyen, J. Hentschke, J. Dedek, D. Prager, and N. Latz. 2005. Epizootiological investigations of parvovirus infections in free-ranging carnivores from Germany. *Journal of Wildlife Diseases* 41: 231-235.

Gabriel, M.W., G.M. Wengert, and R.N. Brown. 2012. Pathogens and parasites of *Martes* species: management and conservation implications. *In* *Biology and Conservation of Martens, Sables, and Fishers: A New Synthesis*, K.B. Aubry, W.J. Zielinski, M.G. Raphael, G. Proulx, and S.W. Buskirk (eds.). Cornell University Press, Ithaca, New York, pp. 138-185.



Methods

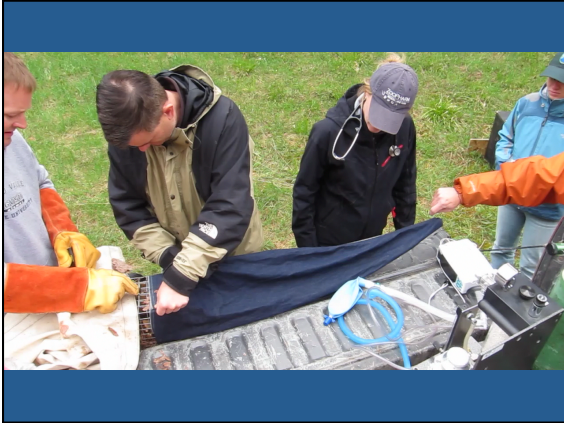
- Tomahawk live-traps
- Trapping summer and winter
- Repeat sampling if re-captured
- Trapper-returned carcasses

A photograph of a tomahawk live-trap set in a snowy forest. The trap is a circular hole in the snow, surrounded by a wire mesh. A small animal is visible inside the trap.

Methods: Sample collection

- Isoflurane anesthesia (Desmarchelier et al., 2007)
- Samples collected:
 - Blood
 - Urine
 - Feces
 - Hair
 - Ear punch biopsy
- Sterile permanent microchip

Two photographs of a marten being handled for sample collection. The top photo shows the marten lying on its back, and the bottom photo shows the marten lying on its side. Both photos show a small white tag on the marten's ear.



Objectives

1. Establish baseline health information including reference intervals for hematologic and biochemical parameters.
 - Cell blood count
 - Blood gas analysis
 - Serum chemistries

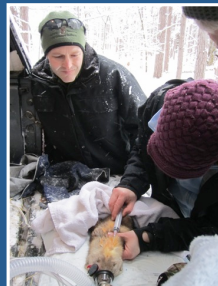
2. Determine pathogen exposure.
3. Assess impact of mercury exposure.
4. Evaluate any differences between the two study sites.
5. Publish data that can be referenced for future management recommendations.

Clinical Pathology

- No published reference intervals for *Martes americana*

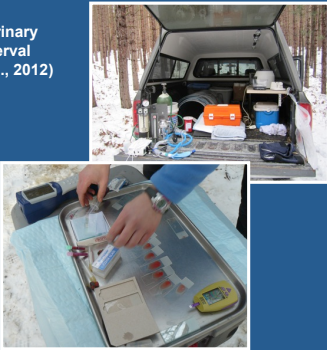
Null Hypothesis:

There will be no difference in hematologic, blood gas, and serum chemistry values between the sexes, age categories, study sites, or collaring status.




Methods

- American Society of Veterinary Pathologists reference interval guidelines (Friedrichs et al., 2012)
- In the field:
 - Blood smear
 - Blood gas analysis
 - Serum chemistries
 - Hematocrit
- In the lab:
 - Cell blood count



Methods

- Reference interval exclusion criteria:
 - Radio-collared
 - Unhealthy on physical exam
 - Sample interference



ANOVA to determine differences between the study sites, sex, age categories in the parameters.

Use correlation and regression to examine influence of variables of interest on each other.

Objectives

1. Establish baseline health information including reference intervals for hematologic and biochemical parameters.
2. Determine pathogen exposure.
 - Toxoplasma gondii*
 - Canine distemper virus
 - Leptospirosis
 - Endoparasitism
3. Assess mercury exposure.
4. Evaluate any differences between the two study sites.
5. Publish data that can be referenced for future management recommendations.

Toxoplasma gondii

- Protozoal parasite with felid as definitive host
- Cause of mortality in other mustelids (Pridham and Belcher, 1958; Cole et al., 2000; Burns et al., 2003b)
- Subclinical effects are important as well (McAllister, 2005; Larkin et al., 2011; Gabriel et al., 2012)



Hypothesis:

Marten in the Manistee NF will have higher seroprevalence of toxoplasmosis compared to the Hiawatha NF due to increased proximity of the Manistee NF to residential areas.



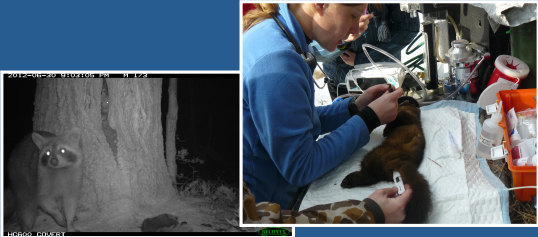
Canine distemper virus

- Morbillivirus, carnivore hosts
- Mustelids are particularly susceptible
- Vaccination strategies employed for black-footed ferret, free-ranging sea otters, Siberian polecat (Williams et al., 1996; Wimsatt et al., 2003; Jessup et al., 2009)



Hypothesis:

Consistent with other species of *Mustelidae*, American marten in both study sites will have a low or zero seroprevalence of canine distemper virus, as infected animals are likely to die.




Leptospirosis

- Rodents are primary hosts
- Possible factor in decline of European mink (Moinet et al., 2010)
- Present in wildlife in Michigan but impact on American marten is unknown (Michigan DNR, 2012)



Hypothesis:

American marten will have a similar seroprevalence to *Leptospira* spp. in both study sites and infected marten have the potential to shed the organism in urine.



Endoparasites

•Parasite fauna of martens in Lower Peninsula is unknown

•Local ecological factors were important in determining parasite prevalence and richness in the Upper Peninsula (Veine-Smith et al., 2011)



Hypothesis:

American marten in the Manistee National Forest will have a different parasite community than those of the Hiawatha National Forest as local ecological factors are different in each site.



Chi-square or t-test to determine differences between sex, age categories, or study site and seropositive titer or presence of a parasite infection.

Prevalence of each parasite may be compared between the study sites using ANOVA.

Objectives

1. Establish baseline health information including reference intervals for hematologic and biochemical parameters.

2. Determine pathogen exposure.


3. Assess mercury exposure.
Mercury concentrations
Selenium concentrations
Nitrogen stable isotope ratio

4. Evaluate any differences between the two study sites.

5. Publish data that can be referenced for future management recommendations.

Mercury in the Great Lakes

- Anthropogenic mercury emissions
- Persistent in the environment
- Bioaccumulates up the food chain
- Fish advisory in place



Great Lakes Mercury Connections


The Extent and Effects of Mercury Pollution in the Great Lakes Region—A Summary

The findings from a binational scientific study

Mercury pollution is a local, regional, and global environmental problem that adversely affects human and pollution have created a substantial recovery challenge for the region.

Mercury, selenium, stable isotopes


- Mercury concentrations vary between tissues (Brookens et al., 2008)
- Selenium reduces toxic effects (Woshner et al., 2011b)
- Isotopic concentrations reflect diet and trophic position (Brookens et al., 2008)



Hypotheses:


Concentration of mercury and selenium in hair will correlate with liver and kidney concentration.

Mercury concentration will be reflective of trophic position as determined by nitrogen stable isotope ratio, $\delta^{15}N$.



Methods

- Mercury & selenium concentrations in hair, liver, kidney
- Nitrogen stable isotope ratio in packed red blood cells, hair, liver



Chi-square will be used to determine differences in concentrations between different tissue types. Correlation can be used to determine relationship between mercury and selenium.

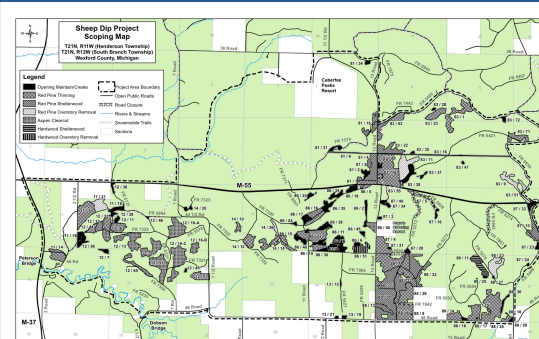
Linear regression to examine relationship between $\delta^{15}N$ and mercury.

Objectives



1. Establish baseline health information including reference intervals for hematologic and biochemical parameters.
2. Determine pathogen exposure.
3. Assess mercury exposure.
4. Evaluate any differences between the two study sites.
5. Publish data that can be referenced for future management recommendations.

Conserving marten through management





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