






Effects of switching to a two-wire hair sampling system for capture-mark-recapture analysis







UF Kaitlin O'Connell, MS Candidate
 University of Tennessee
 Department of Forestry, Wildlife and Fisheries
 November 21st, 2012
 12:20PM PBB 160

Outline

- Capture-Mark-Recapture basics and assumptions
- Non-invasive sampling
- Previous research
- Study area
- Methods
- Results
- Research implication and future directions



Capture-Mark-Recapture (CMR)

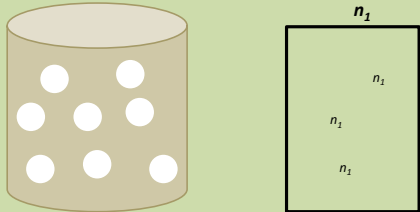
Lincoln – Peterson Model:

$$\hat{N} = \frac{n_1}{\hat{p}}$$

$$\hat{p} = \frac{m_2}{n_2}$$

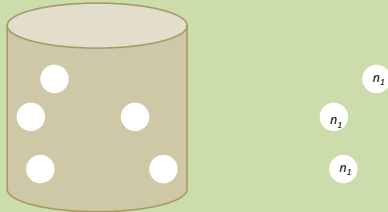
- n_1 = # of animals captured in capture occasion 1
- n_2 = # of animals captured in occasion 2
- m_2 = # of marked animals captured in occasion 2
- p = capture probability

Capture-Mark-Recapture (CMR)

$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$


The diagram shows a cylinder containing 6 white dots. To its right is a rectangular box containing 3 white dots, with the label n_1 above the box.

Capture-Mark-Recapture (CMR)

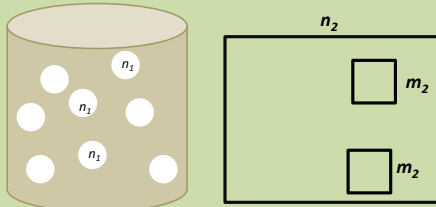
$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$


The diagram shows a cylinder containing 4 white dots. To its right are 3 white dots, with the label n_1 placed above each of them.

Capture-Mark-Recapture (CMR)

$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$

$n_1 = 3$
 $n_2 = 4$
 $m_2 = 2$



The diagram shows a cylinder containing 4 white dots, with 3 of them labeled n_1 . To its right is a rectangular box containing 2 white dots, with the label m_2 next to each dot. Above the box is the label n_2 .

Capture-Mark-Recapture (CMR)

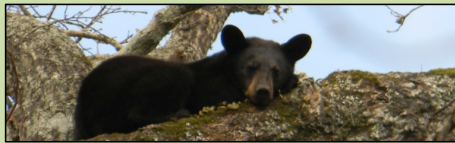
$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$

$n_1 = 3$
 $n_2 = 4$
 $m_2 = 2$

$$\hat{N} = \frac{3}{0.5} = 6 \quad \hat{p} = \frac{2}{4} = 0.5$$

Capture-Mark-Recapture (CMR)

- ✦ Assumptions (Otis et al. 1978):
 1. Population is closed
 2. Marks are not lost
 3. Marks are correctly identified
 4. All individuals have an equal probability of capture and that probability does not change



Capture-Mark-Recapture (CMR)

- 4. All individuals have an equal probability of capture and that probability does not change
 - ✦ **Time** – Capture probabilities vary over time
 - ✦ **Behavioral** – Capture probabilities change due to a behavioral response from being captured (e.g., “trap-happy” or “trap-shy”)
 - ✦ **Heterogeneity** – Capture probabilities differ by individual (e.g., females with cubs vs. females without)
 - ✦ Huggins (1991) – Use of individual covariates
 - ✦ weight, age, or capture frequency
 - ✦ Pledger (2000) – Mixture Models
 - ✦ 2 proportions (π) of the population with different capture probabilities
 - ✦ Hard to catch (π) vs. Easy to catch ($1 - \pi$)

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

Lincoln – Peterson Model:

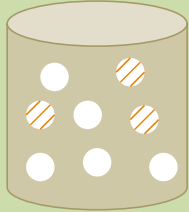
$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$

Capture Probabilities Using Pledger's Mixture Models:

$$\hat{p} = \pi_L * p_L + \pi_H * p_H$$

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

$$\hat{p} = \pi_L * p_L + \pi_H * p_H$$

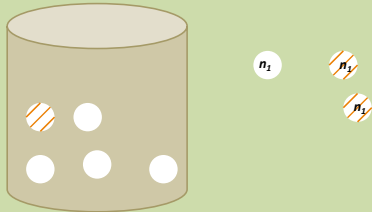
$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$


n_1

- n_1 = # of *animals* captured in capture occasion 1
- n_2 = # of *animals* captured in occasion 2
- m_2 = # of *marked* animals captured in occasion 2
- π_L = Proportion of individuals with a low capture probability
- π_H = Proportion of individuals with a high capture probability

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

$$\hat{p} = \pi_L * p_L + \pi_H * p_H$$

$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$


n_1 n_2 n_2

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

$$\hat{p} = \pi_L * p_L + \pi_H * p_H$$

$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$

The diagram shows a cylinder with 8 balls: 3 with diagonal stripes (marked) and 5 plain white (unmarked). To the right is a 2x2 grid of 4 balls, all with diagonal stripes (marked).

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

$$\hat{p} = \pi_L * p_L + \pi_H * p_H$$

$$\hat{N} = \frac{n_1}{\hat{p}} \quad \hat{p} = \frac{m_2}{n_2}$$

The diagram shows a cylinder with 8 balls: 3 with diagonal stripes (marked) and 5 plain white (unmarked). To the right is a 2x2 grid of 4 balls, all with diagonal stripes (marked).

- $n_1 = 3$
- $n_2 = 4$
- $m_2 = 3$
- $\pi_L = 5/8 = 0.625$ of individuals with a low capture probability
- $\pi_H = 3/8 = 0.375$ of individuals with a high capture probability

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

$$\hat{p} = 0.675 * \frac{1}{4} + 0.325 * \frac{3}{4}$$

$$\hat{p} = 0.437$$

$$\hat{N} = \frac{3}{\hat{p}}$$

$$\hat{N} = \frac{3}{0.437} = 6.3$$

- $n_1 = 3$
- $n_2 = 4$
- $m_2 = 3$
- $\pi_L = 5/8 = 0.675$
- $\pi_H = 3/8 = 0.325$

**Capture-Mark-Recapture
with Heterogeneity Mixtures**

$$\hat{p} = 0.675 * \frac{1}{4} + 0.325 * \frac{3}{4} = 0.437$$

$$\hat{p} = 0.437 \qquad \hat{p} = \frac{3}{4} = 0.75$$

$$\hat{N} = \frac{3}{\hat{p}} \qquad \hat{N} = \frac{3}{\hat{p}}$$

$$\hat{N} = \frac{3}{0.437} = 6.3$$

$$\hat{N} = \frac{3}{0.75} = 4$$

Capture-Mark-Recapture (CMR)

🐾 Capture Probabilities

- 🐾 Heterogeneity – “Bane of CMR’s existence”
 - 🐾 Hard to measure
 - 🐾 Minimize or quantify it as much as possible to get reliable population estimates (Petit & Valiere 2006)
- 🐾 Biased p
 - 🐾 High → Underestimate N
 - 🐾 Low → Overestimate N
- 🐾 Want p as high as possible (>0.20)
 - 🐾 Reduces chance that heterogeneity will go undetected
 - 🐾 Can be low due to trap bias or trapping effort

Non-invasive sampling

🐾 Uses DNA as marker


- 🐾 Collection of hair, feces, or feathers

🐾 No physical handling of the animal

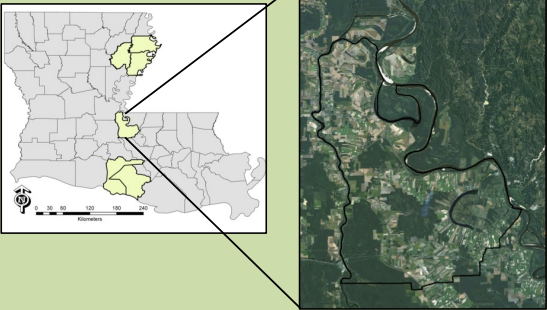
- 🐾 Goal to reduce negative (“trap-shyness”) behavioral biases after first capture

🐾 Increase sampling area and effectiveness

- 🐾 Can cover a larger area and increase the number of “traps” in the study area




**Study Area:
Upper Atchafalaya River Basin**




Sampling Methods

- ✦ Study began in 2007
- ✦ 4 hair sites per home range
- ✦ Grid 1.6 km²
- ✦ Strands of barbed-wire were stretched across 3-4 corner trees
- ✦ Sites were baited and scent lure was hung




Sampling Methods

- ✦ Sites were checked every 7 days for 8 weeks
- ✦ Hair was collected if >5 guard hairs or >20 under-fur hairs.
- ✦ Samples were sent to Wildlife Genetics International for analysis.
- ✦ WGI sub-sampled criteria: 25 samples/week



Methods

2007-2009 (Lowe 2011):



air

Set between 40-60cm

7/11/09 8:18 AM

60 Sec

Change in Methods

2007-2009 (Lowe 2011): 2010-2012:

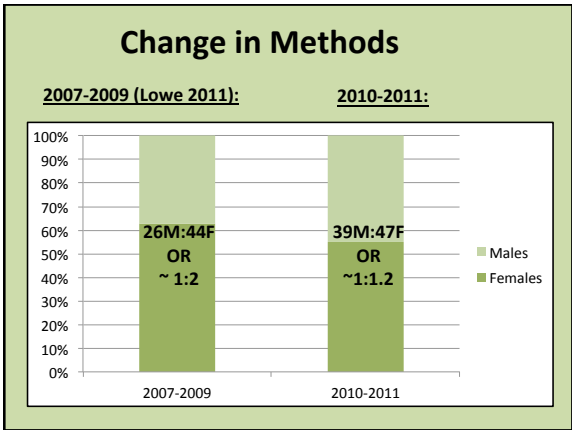


Set between 40-60cm

7/21/11 9:19 AM


60 Sec

Bottom wire: 35-40cm



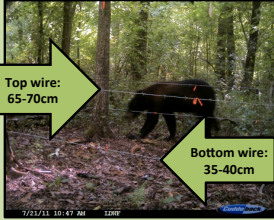
Change in Methods

2007-2009 (Lowe 2011):



Set between
40-60cm

2010-2012:




Top wire:
65-70cm

Bottom wire:
35-40cm

Overall Question:
How does the addition of a second wire affect the estimates of population parameters?

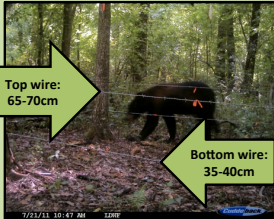
Change in Methods

2007-2009 (Lowe 2011):



Set between
40-60cm

2010-2012:




Top wire:
65-70cm

Bottom wire:
35-40cm

Hypothesis:
Proportion of hard to catch males (π_1) will decrease in year 4 of sampling due to the addition of a second wire.

Methods: Data Analysis

- 🐾 Data from 2007-2010 (2011 excluded)
- 🐾 Program MARK (White and Burnham 1999)
 - 🐾 Closed capture with heterogeneity
 - 🐾 π , ρ , & N
 - 🐾 Constant (.)
 - 🐾 Group effects (g)
 - 🐾 Time dependent (t)
 - 🐾 One vs. Two-wire (Wire)
 - 🐾 One vs. Two-wire for Males (Wire Male)
- 🐾 AIC for Model Selection



Model Set

| Model | AICc | Delta AICc | AICc Weights | Model Likelihood | Num. Par | Deviance |
|---------------------------------------|----------|------------|--------------|------------------|----------|----------|
| {pi(.), p(g), N(t*g)} | 899.0029 | 0 | 0.24011 | 1 | 12 | 675.4288 |
| {pi(wire), p(g), N(t*g)} | 899.1548 | 0.1519 | 0.22254 | 0.9268 | 13 | 673.5403 |
| {pi(wire male), p(g), N(t*g)} | 899.6355 | 0.6326 | 0.175 | 0.7288 | 13 | 674.021 |
| {pi(g), p(g), N(t*g)} | 900.8414 | 1.8385 | 0.09576 | 0.3988 | 13 | 675.2268 |
| {pi(g+wire), p(g), N(t*g)} | 901.0461 | 2.0432 | 0.08644 | 0.36 | 14 | 673.3878 |
| {pi(wire male), p(wire male), N(t*g)} | 901.6381 | 2.6352 | 0.06429 | 0.2678 | 14 | 673.9798 |
| {pi(g+wire), p(g+wire), N(t*g)} | 902.1512 | 3.1483 | 0.04975 | 0.2072 | 15 | 672.4461 |
| {pi(g), p(g+wire), N(t*g)} | 902.6771 | 3.6742 | 0.03824 | 0.1593 | 14 | 675.0189 |
| {pi(g+wire), p(g), N(g*t)} | 904.4021 | 5.3992 | 0.01614 | 0.0672 | 16 | 672.647 |
| {pi(wire), p(t+g), N(t*g)} | 905.9987 | 6.9958 | 0.00727 | 0.0303 | 22 | 661.8763 |
| {pi(wire), p(wire), N(t*g)} | 907.2976 | 8.2947 | 0.0038 | 0.0158 | 13 | 681.683 |
| {pi(g+wire), p(g+wire), N(g*t)} | 911.5275 | 12.5246 | 0.00046 | 0.0019 | 20 | 671.5404 |
| {pi(g), p(g), N(g)} | 913.6904 | 14.6875 | 0.00016 | 0.0007 | 26 | 661.2586 |
| {pi(g), p(g), N(g*wire)} | 916.5476 | 17.5447 | 0.00004 | 0.0002 | 27 | 662.0303 |
| {full} | 918.7587 | 19.7558 | 0.00001 | 0 | 30 | 657.9652 |


Model Set

| Model | AICc | Delta AICc | AICc Weights | Model Likelihood | Num. Par | Deviance |
|-------------------------------|----------|------------|--------------|------------------|----------|----------|
| {pi(.), p(g), N(t*g)} | 899.0029 | 0 | 0.24011 | 1 | 12 | 675.4288 |
| {pi(wire), p(g), N(t*g)} | 899.1548 | 0.1519 | 0.22254 | 0.9268 | 13 | 673.5403 |
| {pi(wire male), p(g), N(t*g)} | 899.6355 | 0.6326 | 0.175 | 0.7288 | 13 | 674.021 |
| {pi(g), p(g), N(t*g)} | 900.8414 | 1.8385 | 0.09576 | 0.3988 | 13 | 675.2268 |
| {pi(g+wire), p(g), N(t*g)} | 901.0461 | 2.0432 | 0.08644 | 0.36 | 14 | 673.3878 |

β (wire), β (wire male), and β (g + wire) did not differ from zero

Research Implications & Future Directions

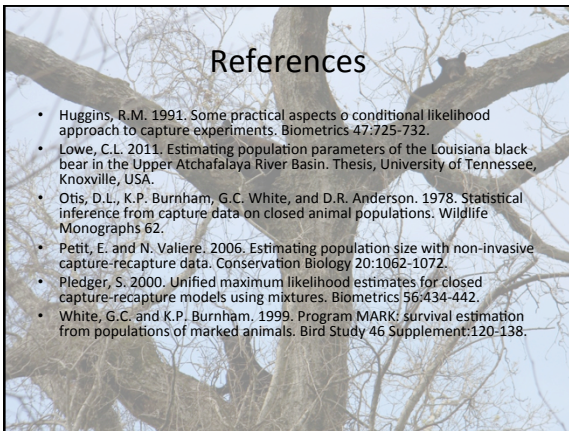
- 🐾 No over-whelming results
- 🐾 Needs more data
 - 🐾 Comparing 3 years vs. 1 year of data
 - 🐾 Increase sub-sample size
 - 🐾 2012 data
- 🐾 Continue with 2-wire system





Acknowledgements

- Committee Members:
 - Dr. Joe Clark, Major Advisor
 - Dr. Frank Van Manen
 - Dr. Arnold Saxton
 - Dr. Ben Fitzgerald
- Funding agencies:
 - USFWS – Debbie Fuller
 - LDWF – Maria Davidson
 - BBCC
 - USGS
 - UTK
 - USACOE
- Many landowners in Pointe Coupee Parish
- "Bear Lab" members
- Technicians: Alex Swaringen, Matt Parker, & Logan Moon



References

- Huggins, R.M. 1991. Some practical aspects o conditional likelihood approach to capture experiments. *Biometrics* 47:725-732.
- Lowe, C.L. 2011. Estimating population parameters of the Louisiana black bear in the Upper Atchafalaya River Basin. Thesis, University of Tennessee, Knoxville, USA.
- Otis, D.L., K.P. Burnham, G.C. White, and D.R. Anderson. 1978. Statistical inference from capture data on closed animal populations. *Wildlife Monographs* 62.
- Petit, E. and N. Valiere. 2006. Estimating population size with non-invasive capture-recapture data. *Conservation Biology* 20:1062-1072.
- Pledger, S. 2000. Unified maximum likelihood estimates for closed capture-recapture models using mixtures. *Biometrics* 56:434-442.
- White, G.C. and K.P. Burnham. 1999. Program MARK: survival estimation from populations of marked animals. *Bird Study* 46 Supplement:120-138.



Photo References

- <http://www.themodernapprentice.com/feathers.htm>
- Goode, M. 2010 <http://fwf.ag.utk.edu/mgray/wfs512/SeminarFA10/Goode.pdf>
- http://nccompart.blogspot.com/2011/01/artistic-discovery-1_21.html
- Carrie Lowe. 2007-2009.