

Impacts of Climate Change and Human Land Use on Southern Appalachian Salamanders





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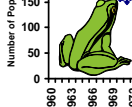
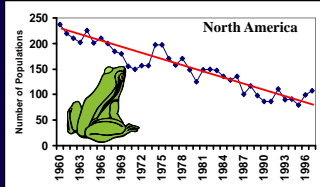



Amphibian Declines

Science
306:1783-1786

IUCN
2008

Science
253:860


32.5% of Amphibians and 46% of salamanders are threatened globally

Why are Amphibian Declining?

- Climate Change
- Habitat Loss
- Diseases & Pathogens
- Pesticides
- Invasive Species
- Overexploitation

Amphibian Declines & Climate Change

Yellowstone NP




Rana luteiventris


PNAS 105:16988-16993

Costa Rica



Atelopus sp.

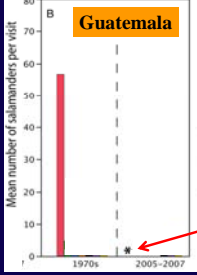
Nature 427:107-109
Nature 439:161-167




Bufo perigrinus

Salamander Declines & Climate Change

Guatemala

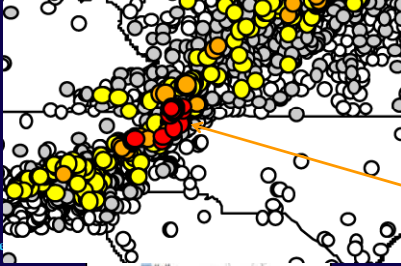


PNAS 106: 3231-3236



Bolitoglossa rostrata

Salamander Declines & Climate Change




Young

Adams 2008

Unique Study Animals

Plethodon welleri








Desmognathus wrighti



Unique Study Animals




Plethodon yanahlossee



Spatial Distributions of Southern Appalachian Salamanders

Previous Research



Dr. James Organ, City College – NY

- Mount Rogers National Recreation Area - VA
- Sampled 1957 - 59 and 1990 - 1991
- Established 14 transects (2950' - 5700')
- No distributional comparisons have been made
- Sites have not been surveyed since 1991

Objectives – Distribution Changes

- 1) Document if salamander distribution changes have occurred since 1950s and 1990s to 2008-10
- 2) Relate potential changes to long term climate change
- 3) Model future spatial shifts due to climate change scenarios



Amphibian Declines & Habitat Loss

Brazil



Physalaemus olfersii

Young et al. 2004

California



Rana draytonii

Conservation Biology 16:1588-1601

Salamander Declines & Habitat Loss

What about the South

Conservation Biology 16:1324-32
Canadian J. Zoology 84:797-807



Conservation Biology 7:363-70
9:983-9
Conservation Biology 21:159-67

Loss of 14 million salamanders
in NC annually due to clear-
cutting

Recovery times of at least 14 years
Forest Ecology and Management 114:245-52

Amphibian Declines & Habitat Loss (Right-of-Ways)

- 2.8 million hectares of ROWs maintained each year
- Mechanical mowing is the most common maintenance method
- Negative impacts to other species of herpetofauna

Gopherus polyphemus



Amphibian Declines & Habitat Loss (Right-of-Ways)

Few studies have examined potential effects on salamanders



Conservation Biology 12:340-52
Yahner et al. 2001
Conservation Biology 21:159-167

Amphibian Declines & ROWs

ROWs are being used for wetland mitigation areas

- Reduce maintenance
- Serve as wetland bank areas



Daniel Boone National Forest - KY
• Over 160 constructed in ROWs

Unique Study Animal



Hemidactylium scutatum

"In Need of Management"



(Redmond & Scott 1996)

Unique Study Animal

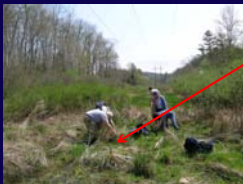


Shallow moss lined temporary pools

Pool breeding is uncommon for
Plethodontid salamanders



4-toed nesting in ROWs



ROW pools with little shade

Forest pools with canopy
cover



Current ROW management



ROW is mowed every 5 years



Do pool temperatures exceed larval thermal limits?

33 - 34°C
Wells 2007

Objectives – ROW management

- 1) Test for differences in larval survival and growth between forest and ROWs pools
- 2) Test the effect of pool distance from the forest edge on larval survival and growth
- 3) Test for differences in larval survival and growth between annual and 5-year mowed ROWs



Amphibian Declines & Diseases

Panama



Atelopus zeteki

Nature 439:143-4

Wyoming




Bufo baxteri

Biological Conservation 110:357-65

Amphibian Declines & Diseases


Ranavirus

Arizona




Ambystoma mavortium

GSMNP



Rana sylvatica



Gray et al. 2009

- Plethodontids are infected
- 81% Prevalence across 10 spp


Gray (unpublished data)

- Ranavirus is pathogenic to Plethodontids


Amphibian Declines - Ranavirus

Gray et al 2009

- Aquatic species have higher infection rates than terrestrial species
- Ranavirus prevalence decreased with elevation




Dr. Organ specimens are housed at Univ. of Michigan Museum of Zoology



- Temperature variation
- UVB
- Downward stream flow
- Aquatic transmission

Objectives – Disease Sampling

- 1) Compare ranavirus prevalence among elevations and species
- 2) Test if distance from stream affects the likelihood of salamander infection
- 3) Determine if ranavirus prevalence has changed since the 1950s



Methods

Climate Change Effects



Study Area

Mount Rogers National Recreation Area



- Highest 2 Mountains in Virginia**
- Mt. Rogers – 5,729'
 - Whitetop Mt – 5,525'
 - Beech Mt - 4,960'
 - Bluff Mt - 4,840'

Grayson, Smyth, & Washington Counties,
Virginia

Study Area

Mount Rogers National Recreation Area



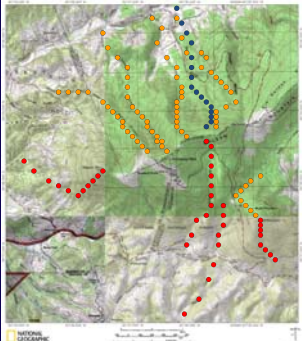
17 salamander species
6 Listed as Special Concern by VADGIF



Methods - Transect Sampling

- Resurvey 10 transects (2008: n=1, 2009: n=5, 2010:n=4)



Red = Southern aspect
Orange/Blue = Northern aspect



2008: Bluff Mt. North
2009: Byars Creek
Whitetop Creek
Dells Branch
Beech Mt.
Daves Ridge
2010: Bills Ridge
Pennington Branch
Bluff Mt. South
Big Branch

Methods - Transect Sampling

Location

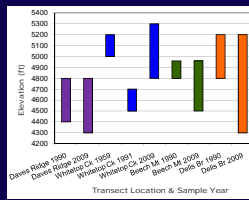
Methods - Transect Sampling





- HOBO Data Loggers every 100'
- Record Temperature and RH every 15 min. from June 1 – Dec 31, 2009

Occupancy Modeling



- Estimate probability of detection among years and sites
- Estimate the likelihood of occurrence using species-specific detection probabilities
- Compare occurrence likelihoods between historic and current surveys using logistic analysis

Climate Modeling

Historic and Current Climate Variables will be compared

Regional Weather Stations can provide historical data:

- Precipitation
- Temperature
- Relative humidity
- Drought index





Ecological Niche Models:

Relate changes in species occurrence to climate variables
Predict future shifts given climate change scenarios

Methods ROW Management



Study Area South Holston Dam

Methods – Nesting Success

1. Locate 4-toed nests & collect nest parameters



Number of eggs



Height above water
Type of moss

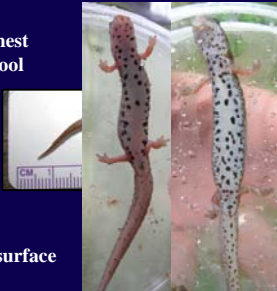
Methods – Nesting Success

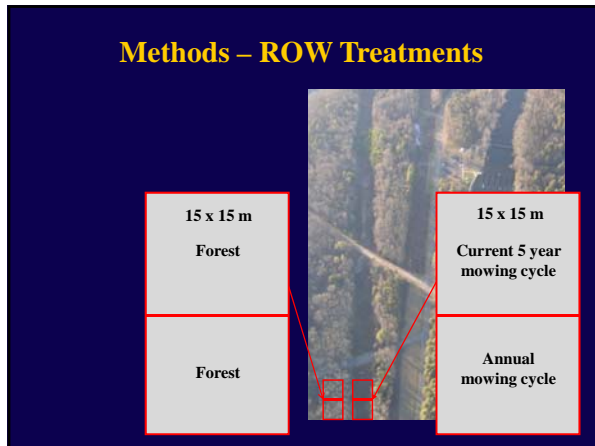
Additional Nest Parameters

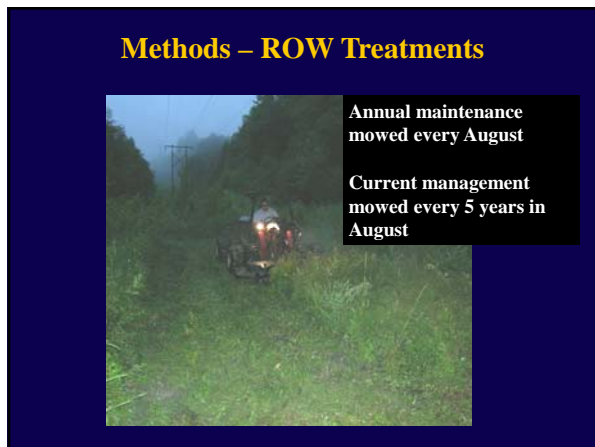
- Nest aspect
- Slope of bank below the nest
- Maximum depth of the pool

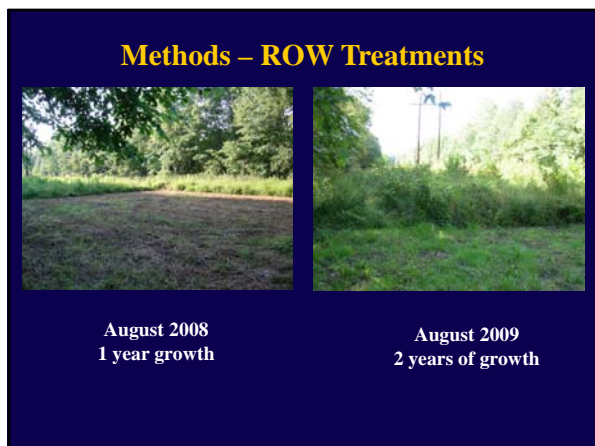
Female Presence

- Determine if nests are joint or single
- Measure SVL & TL
- Photograph female ventral surface for future identification









Methods – ROW Treatments



Methods – ROW Treatments

Place 1-L buckets below nests to catch all hatchlings



Methods – ROW Treatments

- Added to the pool
- 10 larvae
 - 100g leaf litter
 - 2g rabbit chow



Pools searched weekly to determine percent survival



At metamorphosis larvae are collected, weighed, measured (SVL & TL)

Methods – ROW Treatments

Water temperatures and light levels
are measured with HOBO data loggers

33 - 34°C



Pool Productivity

- Quantify phytoplankton and periphyton
- Monitor DO levels for 24 hr periods



Data Collection & Analysis

Response variables

- Growth Rate
- Percent Survival
- Time to Metamorphosis
- Pool Productivity

Effects

- Treatment
 - Annual Mow
 - 5-year Mow
 - Forest
- Distance from the edge

Analysis

- ANOVA

Methods Disease Sampling



Methods – Disease Sampling



Univ. of Michigan
Museum of Zoology

- DNA extracted using Qiagen® kit
- DNA quantified using Qubit flourometer
- Infection tested using conventional and real-time PCR

Data Collection & Analysis

Response variables

- Ranavirus Prevalence

Effects

- Year
- Elevation
- Distance from stream
- Species

Analysis

- Logistic Regression

Special Thanks

- Funding:**
- Virginia Community College System
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 - Tennessee Wildlife Resource Agency



Logistics & Field Work:

- Dr. James Organ
- Mrs. Della Organ
- Gary Poe
- VHCC General Biology Students
- Mt. Rogers National Recreation Area
- VA Dept of Game & Inland Fisheries
- Tennessee Valley Authority

