

Traditional toxicology

Typical protocols: Test model organisms, each species alone, 1-4 days

Taking a tiered approach: Determine LC50s in the lab Maybe manipulate an abiotic variable (pH, temp) Test a few concentrations under field conditions

Determining risk: Which concentrations kill? What's the chance of experiencing those concentrations?



Eco-toxicology: The (not so) New Testament

Single-species tests on model organisms offer a beginning (but are not sufficient) for assessing risk

Organisms always exist as part of a community

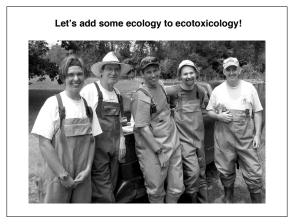
While the goal of traditional tiered research is to include more reality, it ignores the realities of community ecology

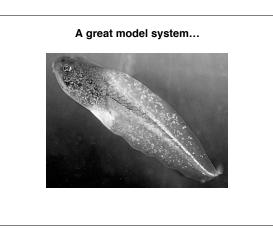
A foreshadowing insight...

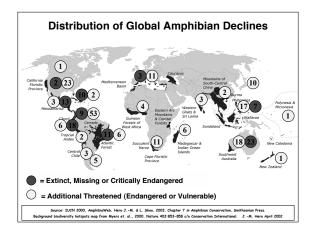
"The chemicals may have been pretested against a few

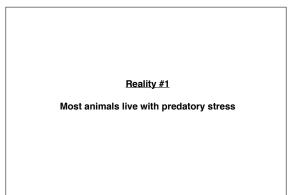
individual species, but not against living communities."

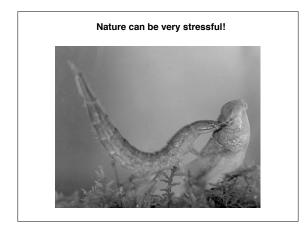
-Rachel Carson (Silent Spring, 1962)



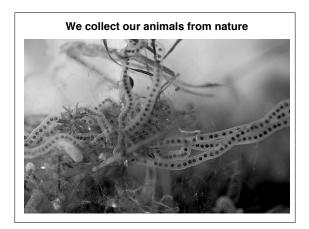


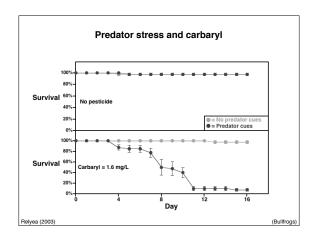


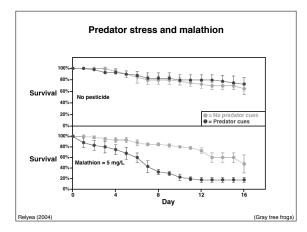


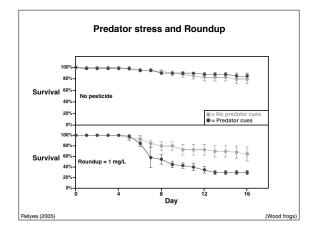


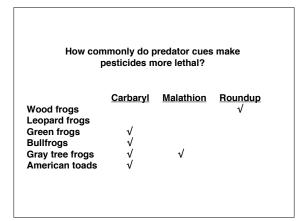


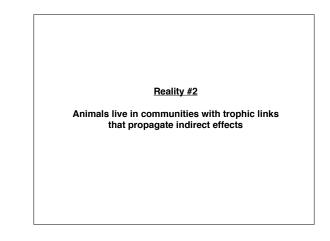


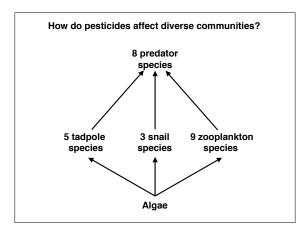


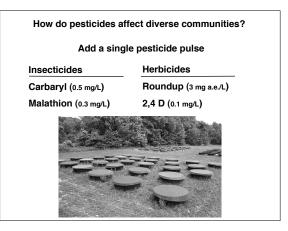


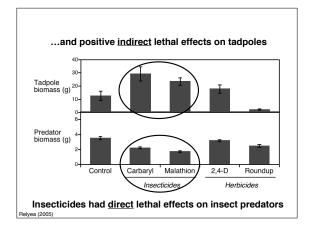


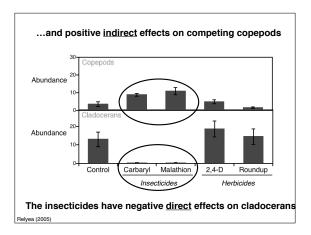


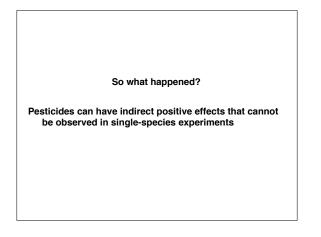












Reality #3

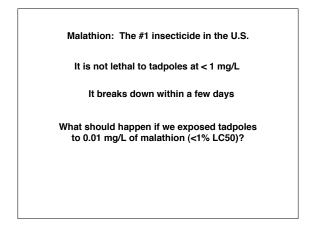
Pesticides concentrations found in nature are typically low

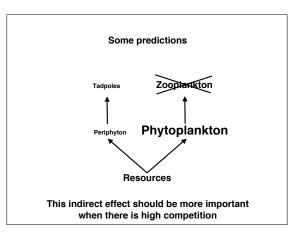
Can low concentrations (1% of LC50) affect amphibians via trophic cascades?

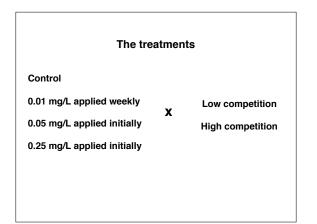
Reality #4

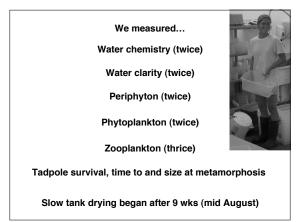
LC50 experiments = constant pesticide concentrations Community experiments = single pulses Nature = multiple pulses

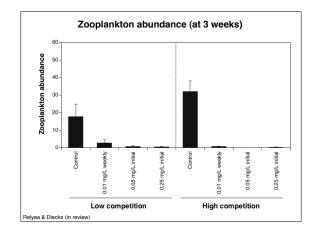
Are several tiny pulses as important as one large pulse?

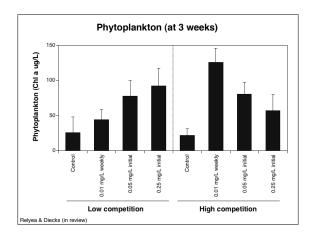


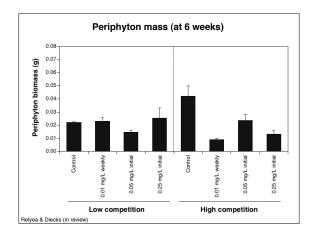


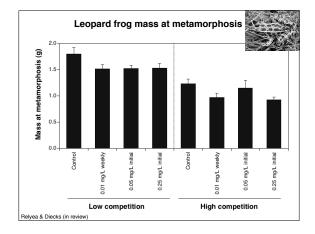


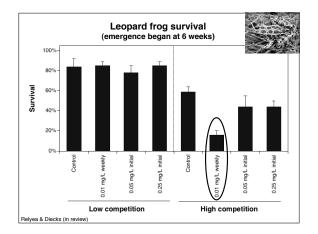


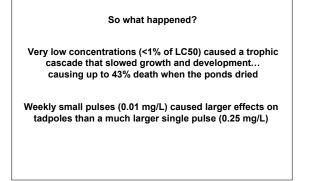


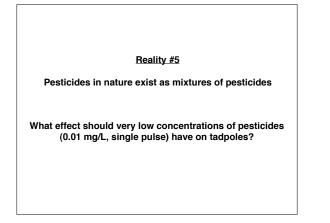




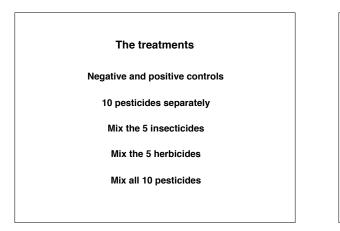


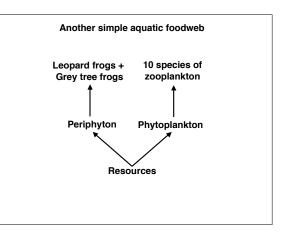


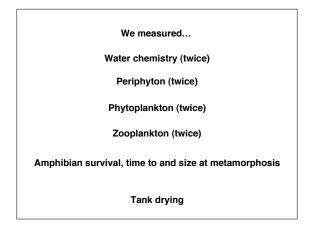


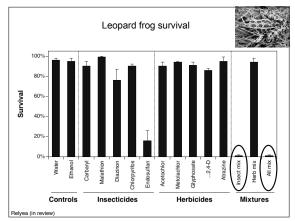


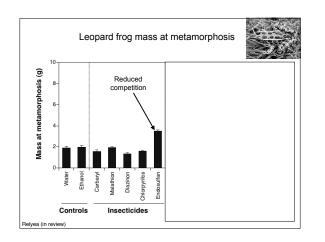
| | EPA drinking | Our expe | Our experiment | |
|--------------|-----------------|----------|----------------|--|
| Insecticides | water standards | Nominal | Actual | |
| Diazinon | Candidate | 10 ppb | 2 ppb | |
| Malathion | No standard | 10 ppb | 6 ppb | |
| Carbaryl | No standard | 10 ppb | 7 ppb | |
| Chlorpyrifos | No standard | 10 ppb | 3 ppb | |
| Endosulfan | No standard | 10 ppb | 6 ppb | |
| Herbicides | | | | |
| Atrazine | 3 ppb | 10 ppb | 6 ppb | |
| Glyphosate | 700 ppb | 10 ppb | 7 ppb | |
| 2,4-D | 70 ppb | 10 ppb | 16 ppb | |
| Acetochlor | Candidate | 10 ppb | 10 ppb | |
| Metolachlor | Candidate | 10 ppb | 7 ppb | |
| | | | | |

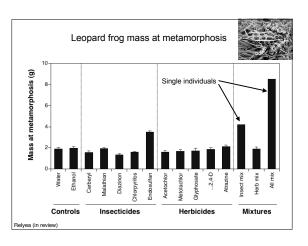


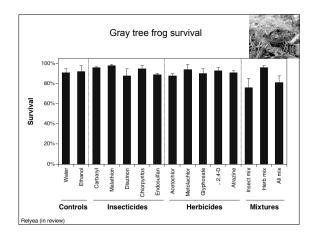


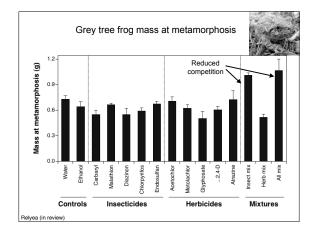












So what happened?

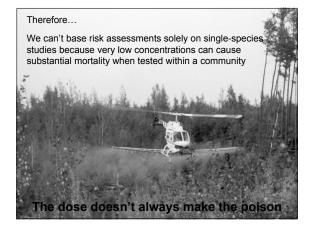
Some single pesticides (e.g., endosulfan) were highly lethal at very low concentrations (~0.01 mg/L)

Mixtures of pesticides at very low concentrations (~0.01 mg/L) can kill up to 99% of some amphibians

The less sensitive tadpole species benefited from reduced competition from the dying species

In Summary...

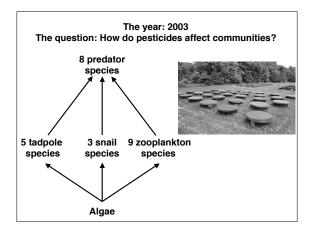
- A large number of pesticides have never been tested on amphibians, yet can be highly lethal (10 ppb)
- Predator stress can make many pesticides up to 46 times more lethal than we ever knew
- Only community-level experiments can detect important indirect effects propagated by other taxa
- Pesticide mixtures can be highly lethal at very low concentrations

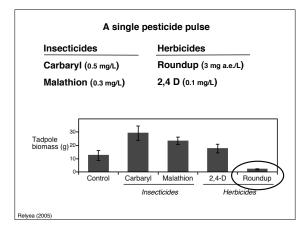


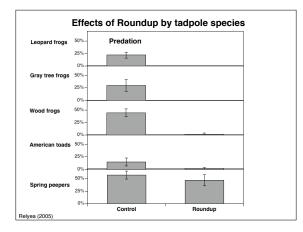
Reality #6

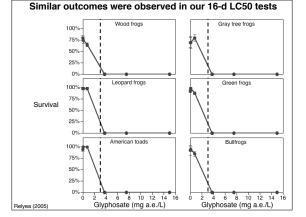
Sometimes LC50 estimates are informative for predicting impacts

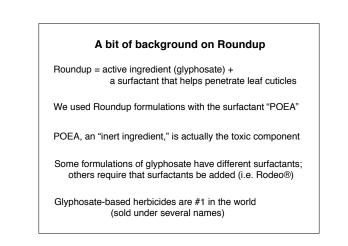
Let's talk about the herbicide Roundup











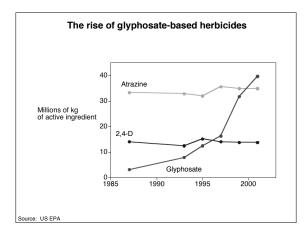
The community experiment was a test of a "worst-case scenario" from a direct overspray of Roundup (3 $_{\tt mg\,asd}$)

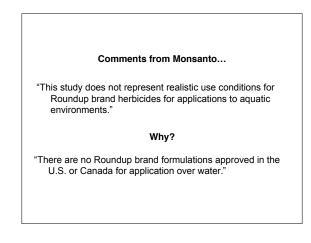
Estimated worst-case scenarios 1.4 mg a.e./L (Canadian gov't)

- 2.7 mg a.e./L (Solomon and Thompson 2003)
- **2.8** mg a.e./L (Giesy et al. 2000)
- 2.9 mg a.e./L (Perkins et al. 2000)
- 7.6 mg a.e./L (Mann and Bidwell 1999)

Observed worst-case scenarios

- 1.2 mg a.e./L (Newton et al. 1994)
- 1.7 mg a.e./L (Monsanto)
- 1.9 mg a.e./L (Thompson et al. 2004)
- 2.8 mg a.e./L (Legris and Couture 1989) 3.1 mg a.e./L (Leville et al. 1993)
- 5.2 mg a.e./L (Edwards et al. 1980)
- _____







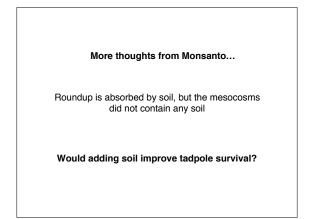
Roundup is not applied to bodies of water?

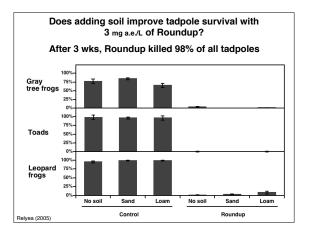
... it depends on your definition of "a body of water"

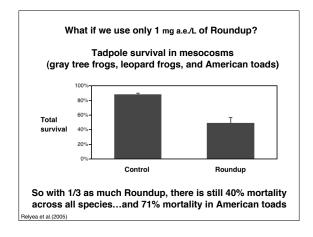
S. 75 (5) of the IPM regulation in Canada:

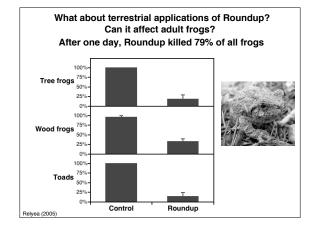
- "The pesticide-free zone is not required along or around a body of water by a person using glyphosate if the body of water is:
 - (a) a temporary free-standing body of water
 - (b) not a wildlife habitat feature
 - (c) not fish bearing, and
 - (d) either smaller than 25 m² or not a wetland"

So Canadian law specifically allows the <u>direct spraying</u> of amphibian habitats with Roundup!













| Species | Mann & Bidwell | Bidwell & Gorrie | Edginton et al | Wojtaszek et al | Howe et al | Relyea | Relyea & |
|---|-------------------|---------------------|-------------------|--------------------|----------------|---------|----------|
| | (2d) | (2d) | et al. (4d) | et al. (4d) | et al. (4d) | (16 d) | (4d) |
| Lymnodynastes dorsalis | 3.0 | | | | | | |
| Heleioporus eyrei | 6.3 | | | | | | |
| Crinia insigniferia | 3.6 | | | | | | |
| Litoria moorei | 2.9 | 11.6 | | | | | |
| Xenopus laevis | | | 0.9-2.1 | | | | - |
| Rana clamitans | | | 1.4-3.5 | 2.7-4.3 | 2.0-7.1 | 1.6 | 1.3 |
| Rana pipiens | | | 1.1-1.8 | 4.3-11.5 | 2.9-6.5 | 1.8 | 1.5 |
| Bufo americanus | | | 1.7-2.9 | | <4-8 | 1.9 | 1.6 |
| Rana sylvatica | | | | | 5.1->8 | 0.4-1.0 | 1.9 |
| Rana catesbeiana | | | | | | 1.6 | 0.7 |
| Hyla versicolor | | | | | | 1.0 | 1.5 |
| Pseudacris crucifer | | | | | | | 0.8 |
| Bufo boreas | | | | | | | 1.9 |
| Rana cascadae | | | | | | | 1.4 |
| Ambystoma maculatum | | | | | | | 3.2 |
| Ambystoma laterale | | | | | | | 3.2 |
| Ambystoma laterale Ambystoma gracile | | | | | | | 3. |
| Notophthalmus viridescens | | | | | | | 2.6 |

