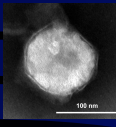
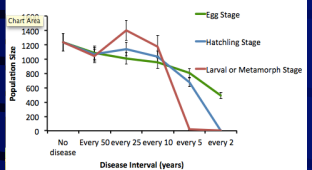

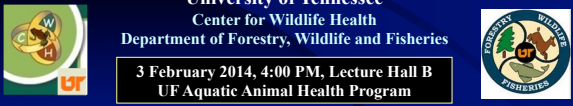


Ranaviruses: Cry Wolf or Real Threat?


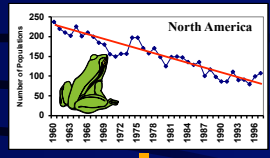






Matthew J. Gray
University of Tennessee
Center for Wildlife Health
Department of Forestry, Wildlife and Fisheries

3 February 2014, 4:00 PM, Lecture Hall B
UF Aquatic Animal Health Program





Presentation Road Map

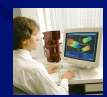





Life History Die-offs

Theory
Monitoring
Simulations

Limited



Ranavirus Characteristics

- dsDNA, 150-280K bp
- 120-300 nm in diameter (3x smaller than bacteria)
- Icosahedral Shape (20)

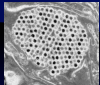
Family: Iridoviridae

Genera: *Iridovirus*, *Chloriridovirus*, *Ranavirus*, *Megalocytivirus*, and *Lymphocystivirus*

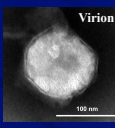
Invertebrates
↓
Ectothermic Vertebrates

Species (6)

- Ambystoma tigrinum* virus (ATV)
- Bohle iridovirus (BIV)
- Frog virus 3 (FV3)
- Epizootic haematopoietic necrosis virus
- European catfish virus
- Santee-Cooper Ranavirus



Paracrystalline Array



Virion

ICTV (2012) Chinchar et al. (2011)

How Does Ranavirus Infect A Host? Routes of Transmission

Indirect Transmission



Water or Sediment
Skin, Gills, Intestines (epithelial cells)
(3 hrs viral transcription)

Direct Contact



One Second Skin Contact

Ingestion



Incidental, Necrophagy, Cannibalism, Predation
(Mortality 2X Faster)

Brunner et al. (2004), Harp & Petranka (2006), Brunner et al. (2007), Hoverman et al. (2010), Robert et al. (2011)

Gross Signs of Infected Amphibians

Edema, Erythema, Hemorrhages, Ulcerations

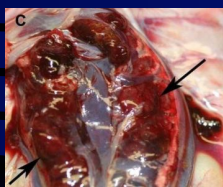


N. Haislip, UT

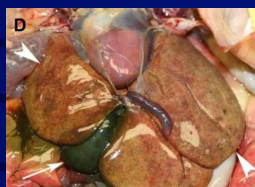
D. Green, USGS

Internal Signs of Ranaviral Disease

Kidney Hemorrhages



Pale and Swollen Liver



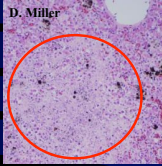
It attacks quickly killing hosts as fast as 3 days!

Hoverman et al. (2011a)

Organ Destruction

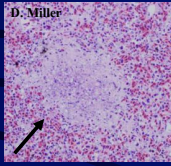
3 Primary Organs: Liver, Spleen, and Kidney

Bollinger et al. (1999)
Miller et al. (2007, 2008, 2011)



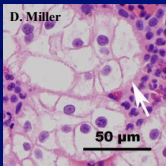
D. Miller

Liver Necrosis



D. Miller

Spleen Necrosis



D. Miller

Kidney Degeneration

Pathogenesis


Target Organ Failure
Heart Failure
Toxicosis, Anemia

?

What if Ranaviruses could Infect Humans?


12 – 32 C
Chinchar (2002)

Monday




Fever

Wednesday




Hands, Feet, Legs Swollen

Friday



Bedridden, Body Enlarged 2X, Lesions, Hemorrhaging Internally and from Orifices

Sunday



Begging Dr. Death (Jack Kevorkian) for a quick end!

No Amphibian Pathogen: Diversity of Gross Signs or Kills as Quickly

There is no Cure!

Mass Mortality Events

Larvae







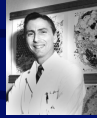
Adults




History of Ranavirus Die-offs

First Isolated:

- Dr. Allan Granoff
- St. Jude Hospital
- Lucke herpesvirus
- Rana pipiens* (1962)



First Large-scale Die-offs:



- Dr. Andrew Cunningham
- Institute of Zoology, ZSL
- Rana temporaria* (1992)



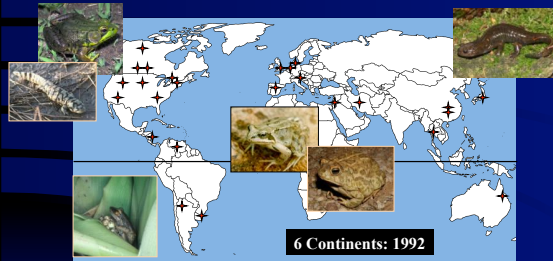
First North American Die-offs:



- Dr. Jim Collins and students
- Arizona State University
- Ambystoma tigrinum stebbinsi* (1985, 1997)



Global Distribution of Ranavirus Cases: Amphibians



6 Continents: 1992

All Latitudes, All Elevations

15 Families: Alytidae, Ranidae, Hylidae, Bufonidae, Leptodactylidae, Dendrobatidae, Discoglossidae, Pipidae, Myobatrachidae, Rhacophoridae, Scaphiopodidae, Ambystomatidae, Salamandridae, Hynobiidae, Cryptobranchidae

>70 Species

Reported Ranavirus Cases in North America: Amphibians



Uncommon

>30 States & 5 Provinces;
46 Species

Families

- Bufonidae
- Hylidae
- Ranidae
- Scaphiopodidae
- Ambystomatidae
- Cryptobranchidae
- Plethodontidae
- Salamandridae



Lithobates sylvaticus

viruses
ISSN 1999-4915

Ecopathology of Ranaviruses Infecting Amphibians
Debra Miller ^{1,2*}, Matthew Gray ¹ and Andrew Storfer ¹ 2011

Case Example

Re-occurring Die-offs

Jamie Barichivich (USGS) and Megan Todd-Thompson (UT)

May 1999, 2000, 2009, 2012

GSMNP, Cades Cove
Gourley Pond

Spotted & Marbled Salamander, Wood Frog,
Spring Peeper, Southeastern Chorus Frog

Green et al. (2002)

A. Cressler, USGS; M. Niemiller, UT; A. Cressler, USGS; B. Green, USGS

Cases of FV3-like Ranaviral Disease in Reptiles

(Westhouse et al. 1996; Marschang et al. 1999, 2005; Hyatt et al. 2002; DeVoe et al. 2004; Huang et al. 2009; Allender et al. 2006, 2011; Johnson et al. 2007, 2008, 2011)

Gopherus polyphemus, Testudo hermanni, Terrapene carolina carolina, Trionyx sinensis, Uroplatus fimbriatus, and Chondropython viridis

Over >95% homology with 1000-bp region of FV3 MCP

At least 20 reptile species

Marschang (2011)

viruses ISSN 1999-0112

What about other Ectothermic Vertebrates?

13 February 2012

The Washington Post Make us your start page

POSTLOCAL

North Branch Stream Valley State Park

Deadly virus hits turtles, tadpoles in Montgomery County

26 of 31 Box Turtles Die from Ranaviral Disease

2008 – 2011

Larval anurans and salamanders dead too

Farnsworth and Seigel (2013)

View Photo Gallery — Biologists say an alarming number of turtles rescued from the path of the Intercountry Connector's construction have died of a virus they fear could devastate Maryland's ecosystem.

Cases of Ranaviral Disease in Fishes



Ictalurus melas, *I. nebulosa*, *Silurus glanis*, *Psetta maxima*, *Sander lucioperca*, *Perca fluviatilis*, *P. flavescens*, *Oncorhynchus mykiss*, *Pomoxis nigromaculatus*, *Gambusia affinis*, *Epinephelus tauvina*

EHN, ECV

At least 30 fish species

LMBV, SGIV

Iridovirus infections in finfish – critical review with emphasis on ranaviruses

Journal of Fish Diseases
33:95-122

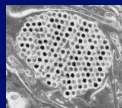
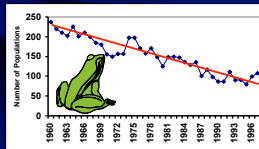
R J Whittington, J A Becker and M M Dennis

World Organisation for Animal Health (OIE) Reference Laboratory for EHN Virus, Faculty of Veterinary Science, The University of Sydney, Sydney, Australia

Are Ranaviruses Capable of Causing Local Extirpations and Species Declines?



Muths et al. (2006)



Collins & Crump (2009)

Traditional Theory (Anderson and May 1979)



Extirpation is possible if:

Frequency Dependent

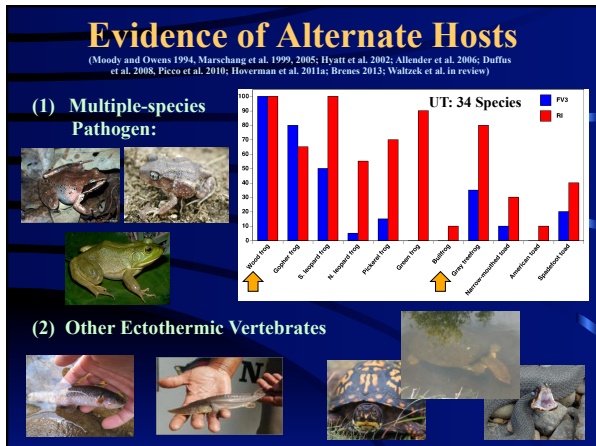
(1) Multiple Host Species Where Susceptibility Differs

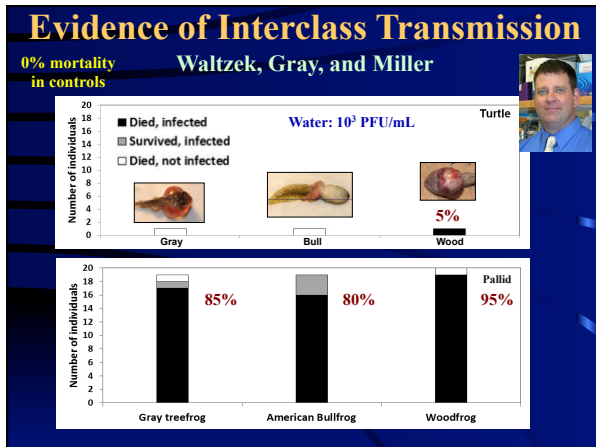
Is at least one of these conditions satisfied in the ranavirus-host system?

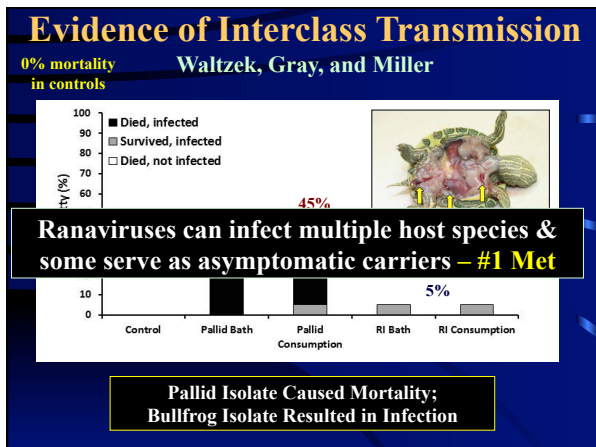
- Survive Outside Host

(3) Clustering of Individuals

- Sexually transmitted disease







Evidence of Environmental Persistence

(1) EHNVP Persistence (Langdon 1989)

Ranaviruses can remain viable outside the host for considerable duration (**permanent wetlands at colder temperatures**). – #2 Met

(2) FV3, FV3-like (Nazir et al. 2012)

20 C = •PW (unsterile): **22-34 d** •Soil: **13-22 d**
 4 C = •PW (unsterile): **58-72 d** •Soil: **30-48 d**

(T-90 Values)

Evidence of Frequency Dependent Transmission

(1) Breeding (Brunner et al. 2004)

Frequency dependent transmission is possible in larval and adult age classes – #3 Likely.

(2) Larval Clustering

- Increase Contact Rates
- Vegetation Reduction

(Greer et al. 2008)

Local Extirpations and Declines?

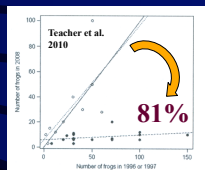
YES, all 3 characteristics met in the Ranavirus-Host System

Evidence of Local Extinction



Dr. Amber Teacher
Southeastern England
1996/97 and 2008

Animal Conservation
13:514-522




Ranavirus (+) populations
81% Median Reduction



Larger Populations
Greatest Proportional Declines


Evidence of Local Extinction



Dr. Jim Petranka
Tulula Wetland Complex, NC
1998-2006


Biological Conservation
138:371-380
Wetlands
23:278-290

Recruitment at most wetlands failed due to **ranavirus**




Persistence Possible from **Source Populations**

Rescue Effect



Uncommon Species?



Should we be concerned with conserving a few uncommon species?

Commonality of Being Uncommon

Southeastern United States


Federally Listed: *Rana sevens,* *Ambystoma cingulatum,*
Phaeognathus hubrichti, *Ambystoma bishopi*

Species of Concern: 113 Species and 25 Genera Total 50% U.S.

- 1) Alabama = 14 species (11 genera)
- 2) Arkansas = 25 species (12 genera)
- 3) Florida = 19 species (12 genera)
- 4) Georgia = 22 species (15 genera)
- 5) Kentucky = 22 species (11 genera)
- 6) Louisiana = 15 species (10 genera)


RISK?
Testing Susceptibility for
Uncommon Species



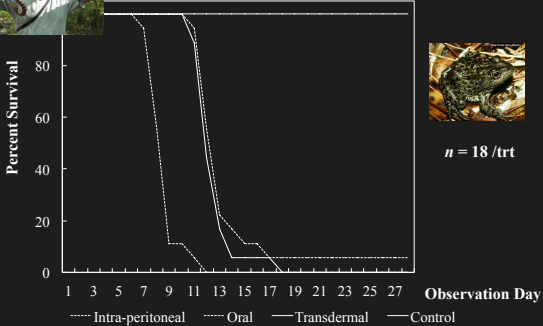


Evidence of Rare Species Effects

Sutton et al. (in review)
Endangered Dusky Gopher Frog




Animal Conservation




n = 18 /trt

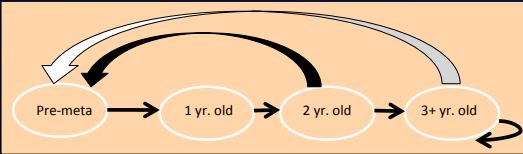
Wood Frog Population Data




Drs. Keith Berven and Elizabeth Harper
Berven (1990), Harper et al. (2008)


7-year: Beltsville, Maryland



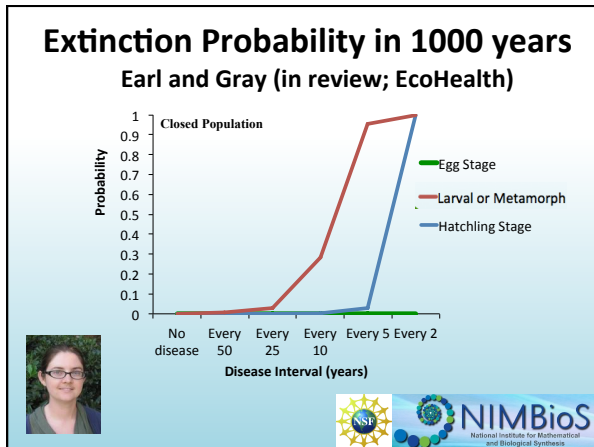


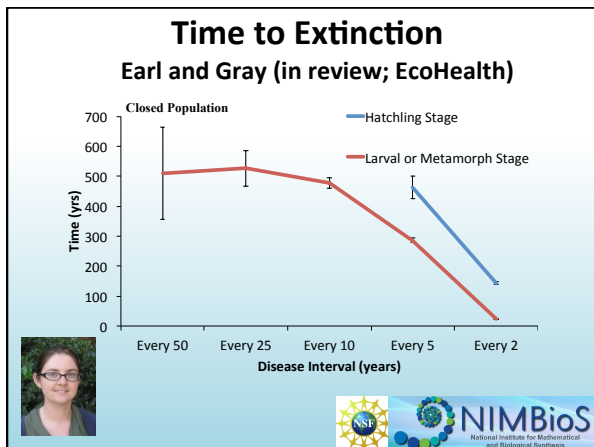


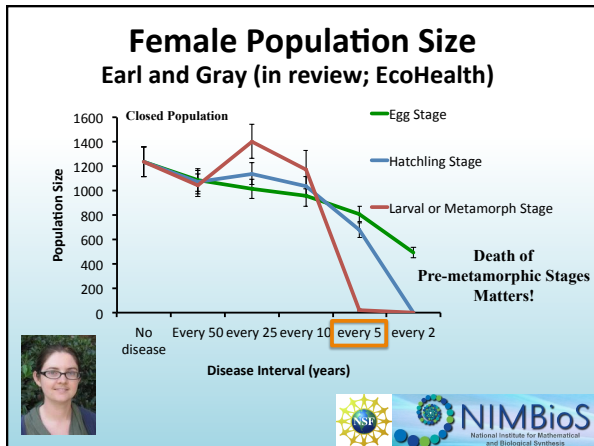
- Stage-structured discrete-time matrix model (K = 1500 females)
- Simulated Exposure for each Stage (egg, hatchling, larva, meta)
- Exposure Interval: **50, 25, 10, 5, and 2 years**
- Demographic and ranavirus survival probabilities multiplied
- 1000 Simulations (one-year steps)
- Built in stochasticity in the model
- Closed Population



J. Earl







Ranavirus Landscape Prevalence

n = 40 ponds
2 years, 4 seasons

Tennessee Ponds
Hoverman et al. (2011b)

Ranaviruses are common hence have the potential to have landscape scale impacts.

Ranavirus Distribution: 83% of Ponds Sampled

Hotspots: ≥40% in 15 out of 40 Ponds Sampled

- 85% Infection
- 100% Infection (morbid and dead)

EcoHEALTH
EcoHealth 9:36-48

Global Trade of Ranavirus Hosts

Kristine Smith, DVM

From 2000-2006, the U.S. imported >1.5 billion individual animals (fish & wildlife; Smith et al. 2009)

- 90% fish, 2% amphibians, 1% reptiles
- 25 million live amphibians imported to U.S./year

Ranavirus Positive

- Hong Kong = 89%
- Dominican Republic = 70%
- Madagascar = 57%

Smith et al. (unpubl. data)

EcoHealth Alliance



Cry Wolf or Valid Risk?



Should we be Concerned?

- Ranavirus Die-offs have Global Distribution
- Ranavirus Prevalence can be High
- Ranaviruses Infect Multiple Ectothermic Vertebrate Species with Different Susceptibilities
- Interclass Transmission is Possible – Abundant Reservoirs
- Ranavirus Persistence is Long
- High Transmission: Breeding and for Schooling Spp.

Epidemiological Theory AND Initial Simulations Support the Premise that Ranaviruses Could Cause Local Population Extirpations and Contribute to Species Declines

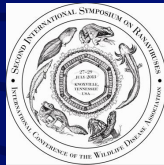
More Research: Species Susceptibility, Community Composition, More Complex Simulations (dispersal, transmission), Long-Term Monitoring

Global Ranavirus Consortium

<http://fwf.ag.utk.edu/mgray/ranavirus/ranavirus.htm>



Symposia
 Discussion Groups
 Website
 Reporting System
 Outreach Resources
 Springer eBook



The goal of the GRC is to facilitate communication and collaboration among scientists and veterinarians conducting research on ranaviruses and diagnosing cases of ranaviral disease

Bylaws Approved

GRC@LISTSERV.UTK.EDU

Membership in 2014

Global Ranavirus Consortium

Executive Committee and Board

 Director Matthew J. Gray, Ph.D. University of Tennessee	 Associate Director Jesse L. Brunner, Ph.D. Washington State University	 Secretary/Treasurer Amanda L. J. Duffus Ph.D. Gordon State College
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