

Pathogenesis and Immuno-control of Singapore Grouper Iridovirus (SGIV)

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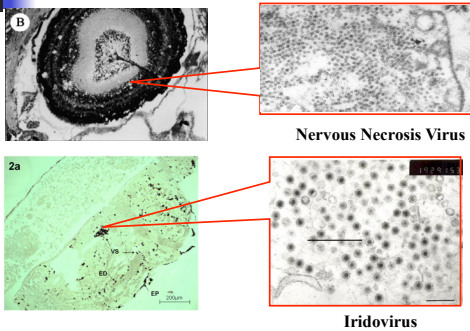
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Groupers are major high-value species being cultured in China and Southeast Asia countries. They are particular popular for the live fish market mainly in Hong Kong, Taiwan, main land of China and SE countries



2

The cultured grouper survival rate is very low under artificial conditions (<40%)
Major problems: Nodavirus and iridovirus outbreaks

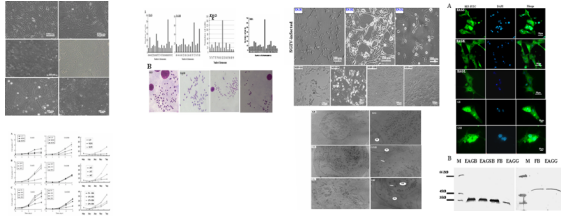


Nervous Necrosis Virus

Iridovirus

3

(I) 9 New fish cell lines susceptible to viruses established: 8 from grouper, 1 from flounder

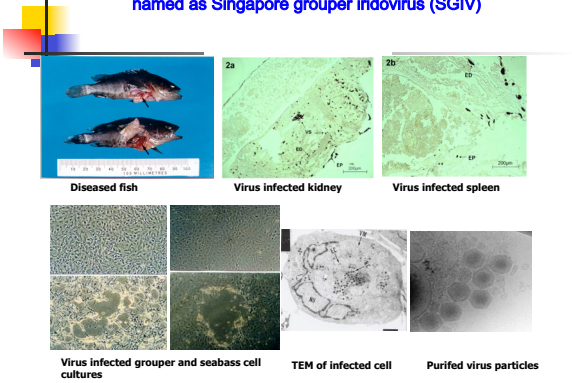


Provide cell culture platform for marine fish virus isolation, pathogenesis and vaccine development

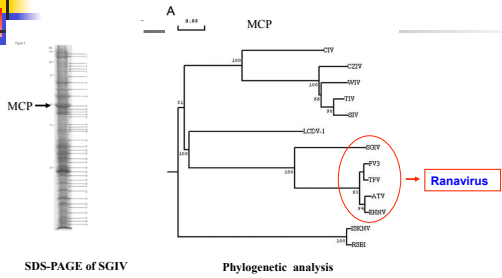
Qin et al., J. Virol Meth, 2006.
 Huang et al., Aquaculture, 2009; 2010.
 Ouyang et al., J. Fish Biol., 2010.
 Gong et al., J. Fish Dis., 2010

4

(II) A novel pathogenic iridovirus isolated from diseased grouper in Singapore, named as Singapore grouper iridovirus (SGIV)



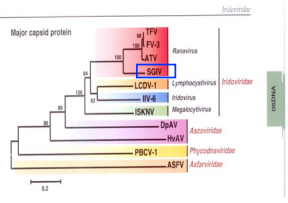
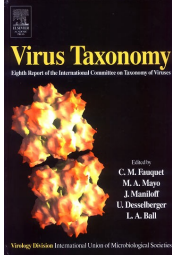
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Qin et al. Dis. Aquat Org., 2003, 53: 1-9.
 Qin et al. J. Virol Meth., 2004, 117: 123-128.

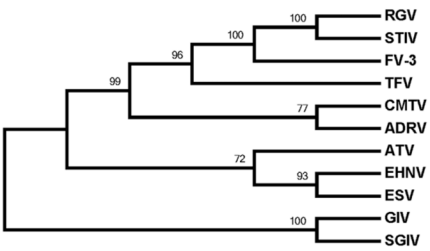
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SGIV has been listed as a tentative new species of genus *ranavirus*, family *iridoviridae*



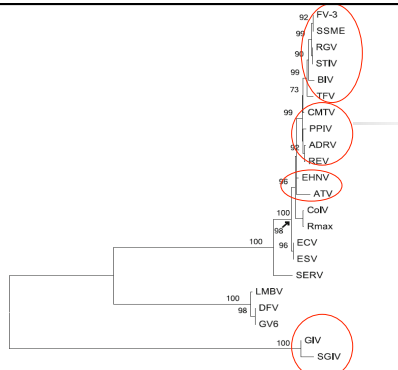
Chinchar et al. 2005. In *Virus Taxonomy*, p161.

7



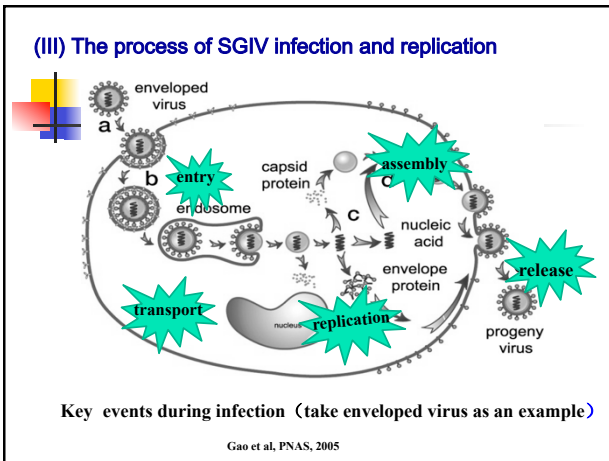
The evolutionary relationships among the 11 fully sequenced ranaviruses, based on aligned deduced amino acid sequences of 26 conserved iridovirus genes (Jancovich et al., 2015).

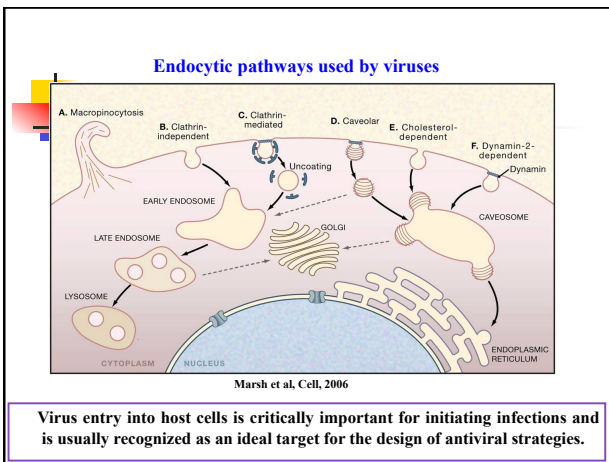
8

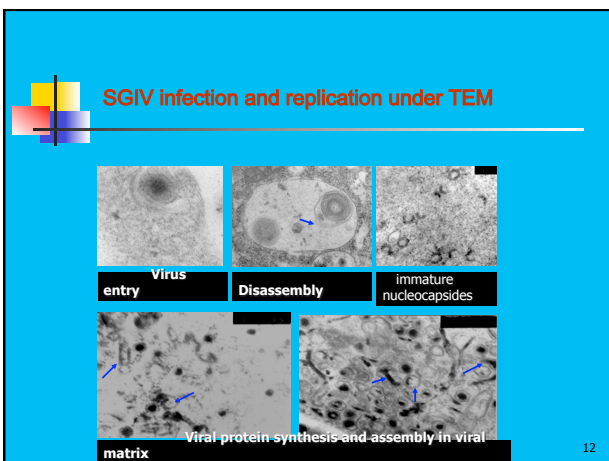


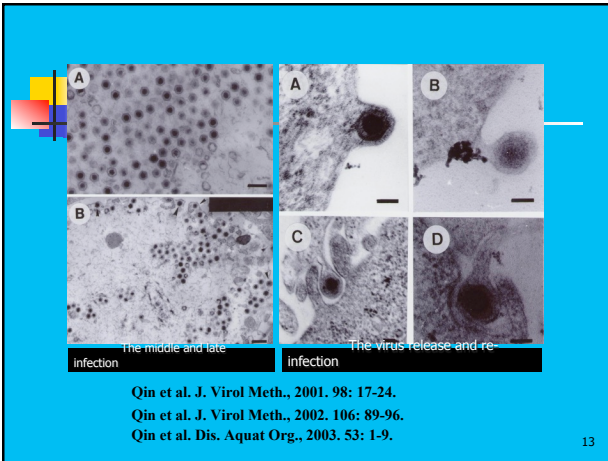
The evolutionary relationships among the 22 ranaviruses, based on full length nucleotide (nt) sequences of major capsid protein (MCP) genes (Jancovich et al., 2015). Ranaviruses could be classified into 4 groups, and group of SGIV & GIV is distinctively different from other 3 groups.

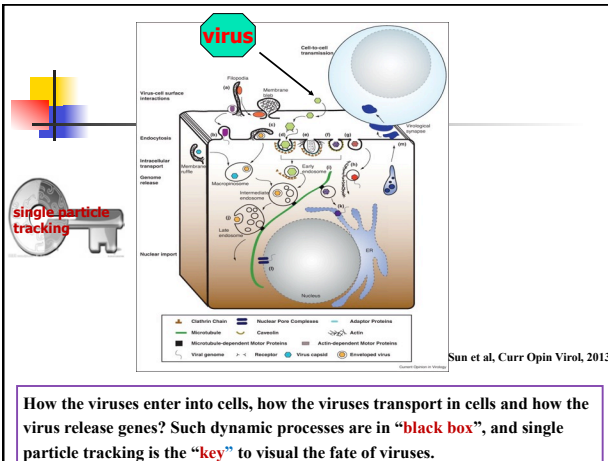
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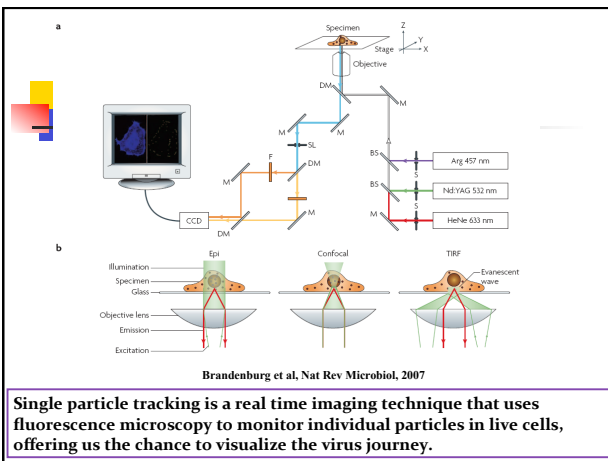


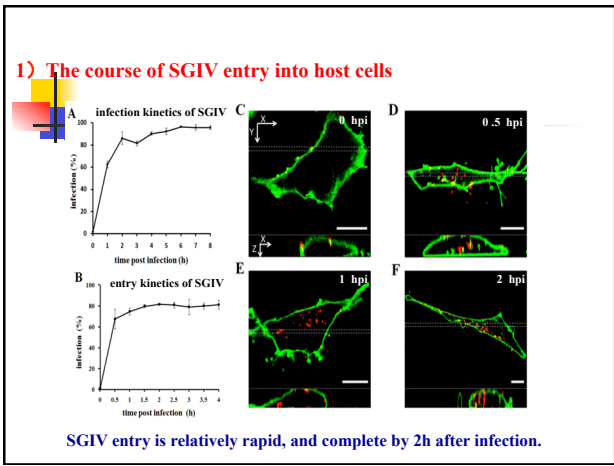


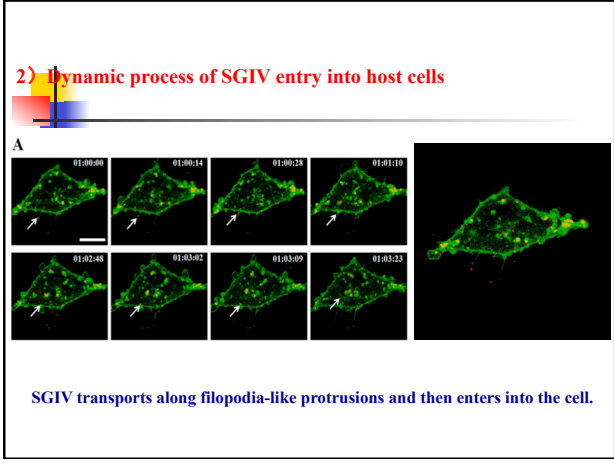


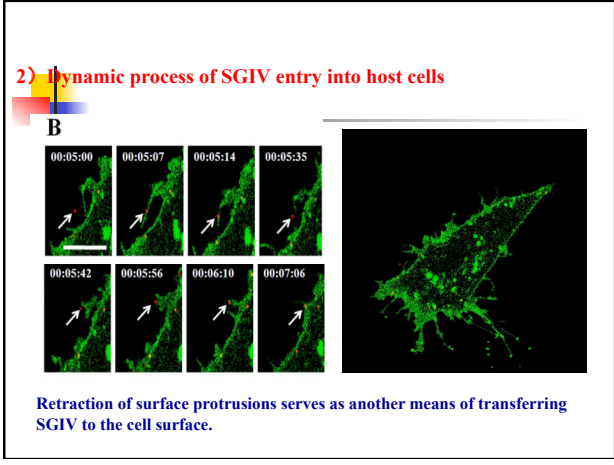




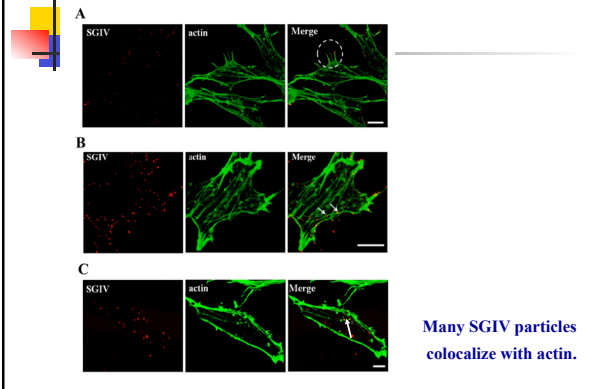




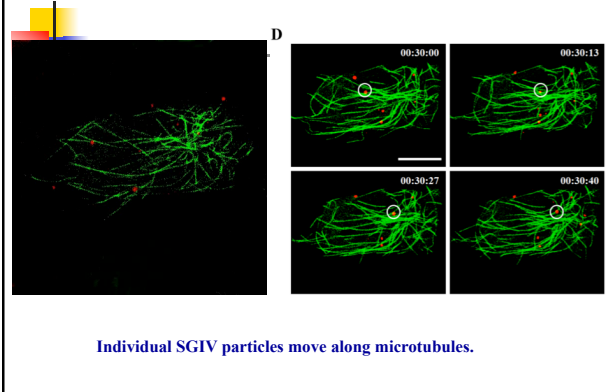




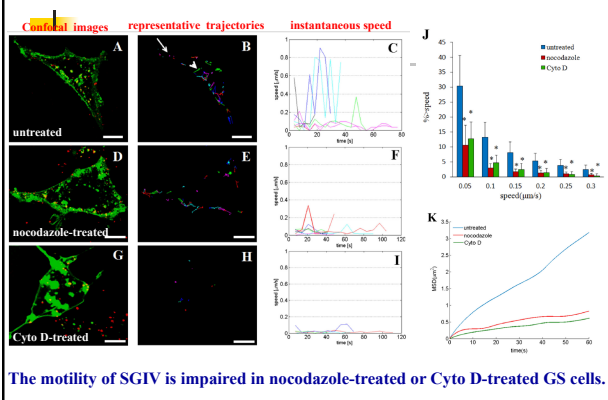
3) The motility of SGIV is dependent on the cytoskeleton



3) The motility of SGIV is dependent on the cytoskeleton

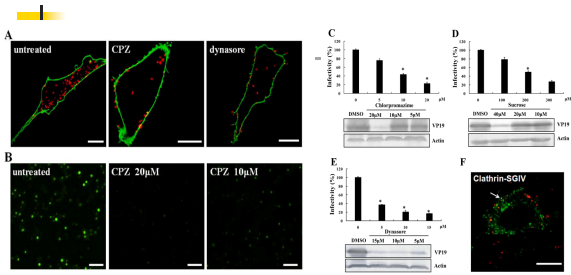


3) The motility of SGIV is dependent on the cytoskeleton



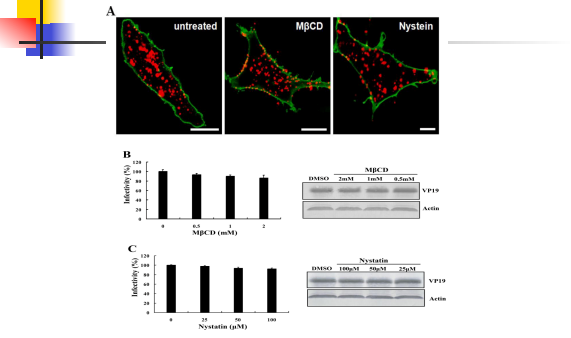
The motility of SGIV is impaired in nocodazole-treated or Cyto D-treated GS cells.

4) The endocytic pathways of SGIV entry



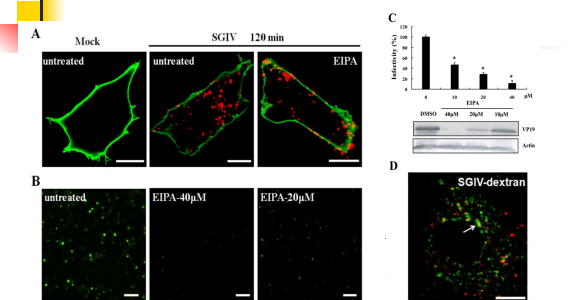
Dynamin-dependent and clathrin-mediated endocytosis is involved in SGIV entry.

4) The endocytic pathways of SGIV entry



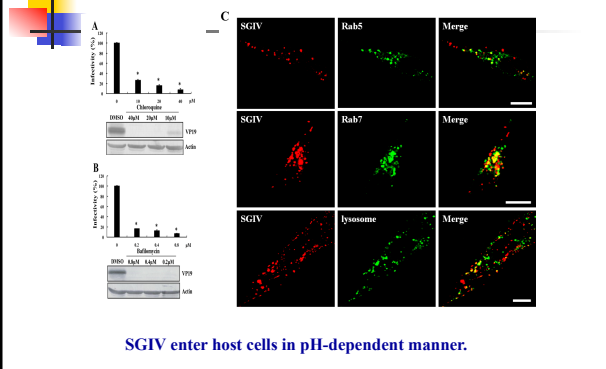
Entry of SGIV is independent of lipid raft/ caveolin-dependent endocytosis.

4) The endocytic pathways of SGIV entry

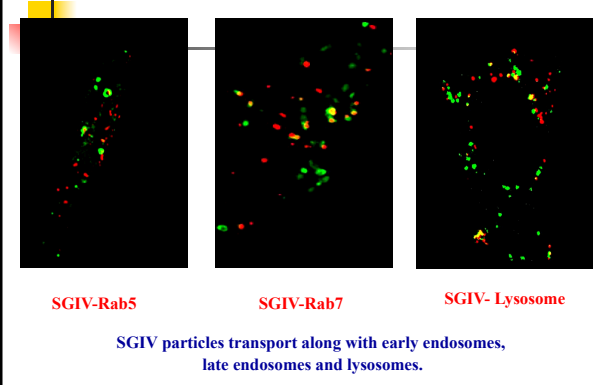


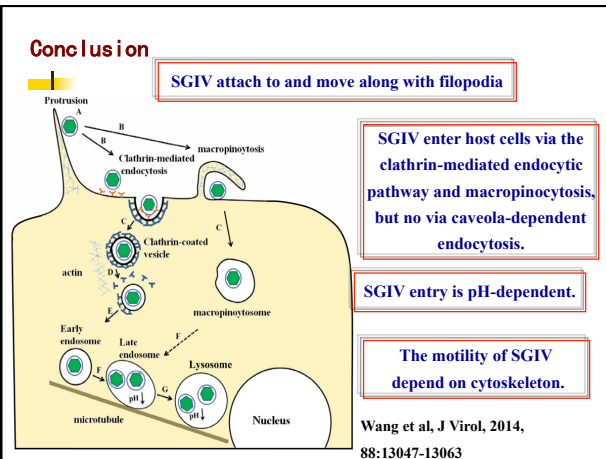
SGIV also uses micropinocytosis to enter GS cells.

4) The endocytic pathways of SGIV entry

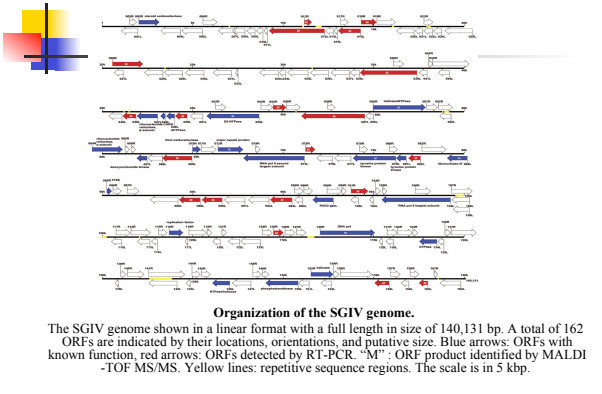


3) the pathways of SGIV entry

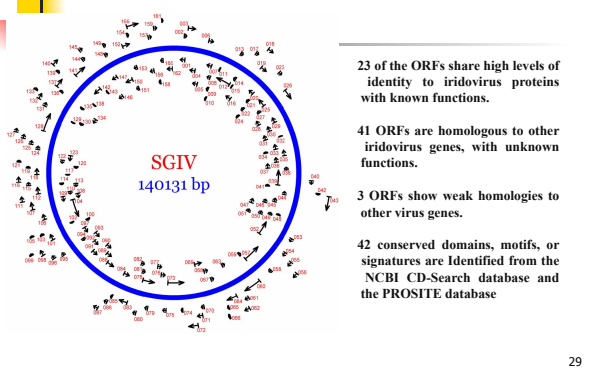




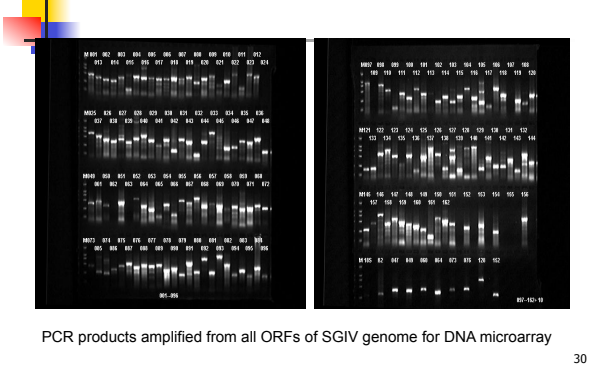
(IV) Genomics of SGIV



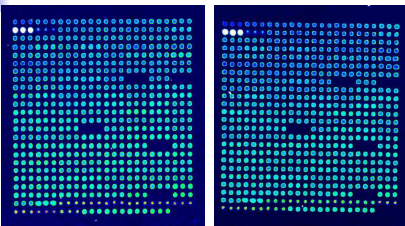
Circular representation of SGIV genome



(V) Transcriptome of SGIV

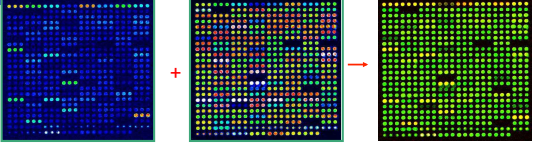


DNA Microarrays containing SGIV all 162 ORFs (The first chip and the last chip)



31

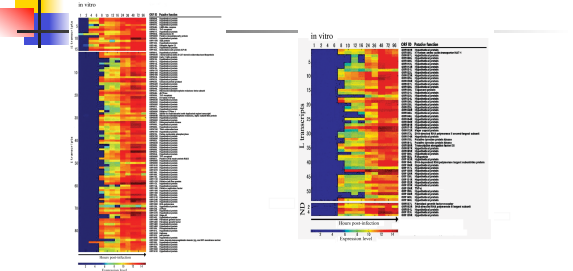
Gene transcription and expression of SGIV in virus-infected cells (post infection of 48 h)



Mock-infected control SGIV-infected GS cells Overlay of infected signals with control

32

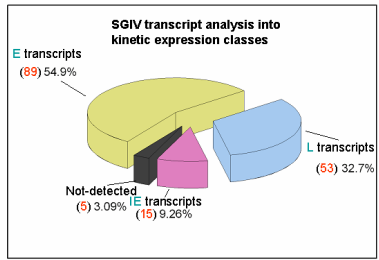
Transcription profiles of SGIV in virus-infected grouper spleen cells invitro



Columns represent separate time points, and each row indicates the transcriptional profile of a single ORF during viral infection.

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Temporal expression kinetics of SGIV transcripts

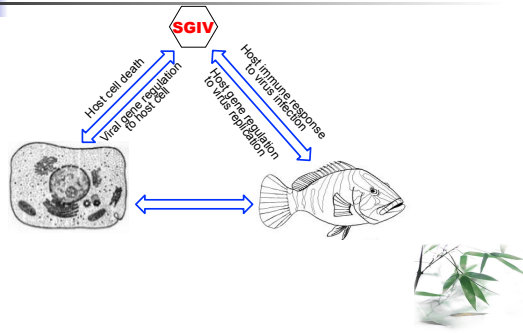


15 immediate early (IE), 89 early (E) and 53 late (L) genes were classified.

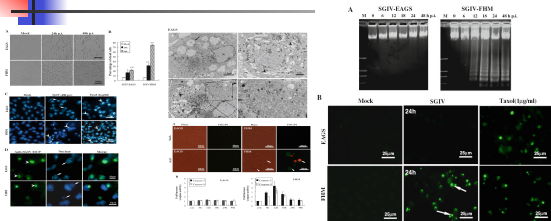
Teng et al., 2008. Virology, 377: 39-48.

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(VI) SGIV pathogenesis: interactions between virus and host



1. SGIV infection induced a new programmed cell death-parapoptosis in grouper host cells



Non-apoptotic bodies, caspase 3, 8,9 not activated, cytoplasmic vacuoles, distended endoplasmic reticulum (ER), large cluster of swollen mitochondria. No DNA ladder, TUNEL negative.

(Huang et al., 2011a Apoptosis)

SGIV infection activated MAPK pathways, ERK and JNK signaling molecules involved in virus replication and virus induced cell death. Inhibition of ERK and JNK activities could delay the CPE and reduce the virus production.

ERK磷酸化检测及其抑制剂对病毒滴度影响

JNK和p38MAPK磷酸化检测及其抑制剂对病毒滴度影响

(Huang et al., 2011b Apoptosis) 37

2. SGIV encoded two IE gene (ICP18, ICP46) could regulate host cell cycle, promote cell growth, contribute virus replication.

SGIV ICP18磷酸化分析

SGIV ICP18磷酸化检测

SGIV ICP18磷酸化分析

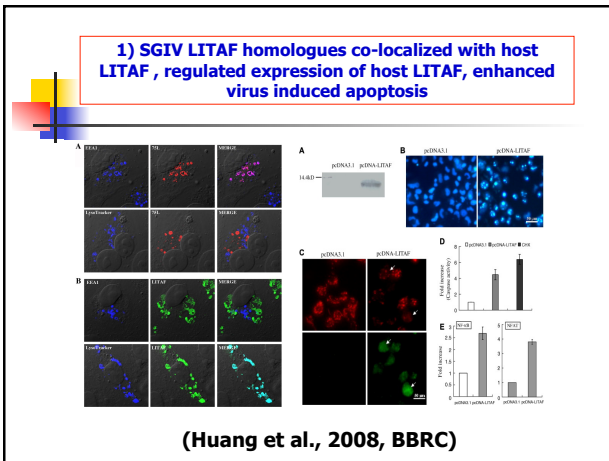
SGIV ICP46磷酸化分析

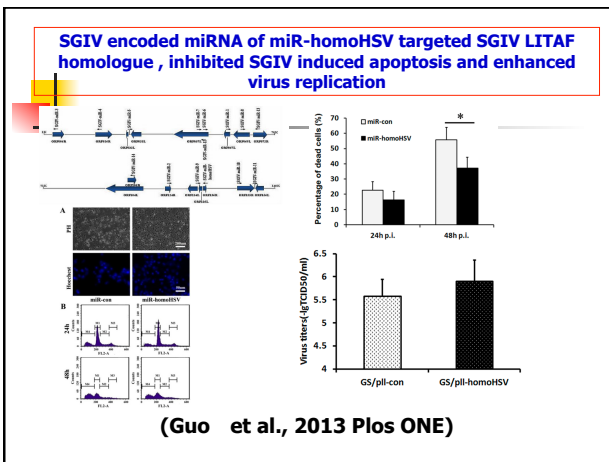
SGIV ICP46磷酸化检测

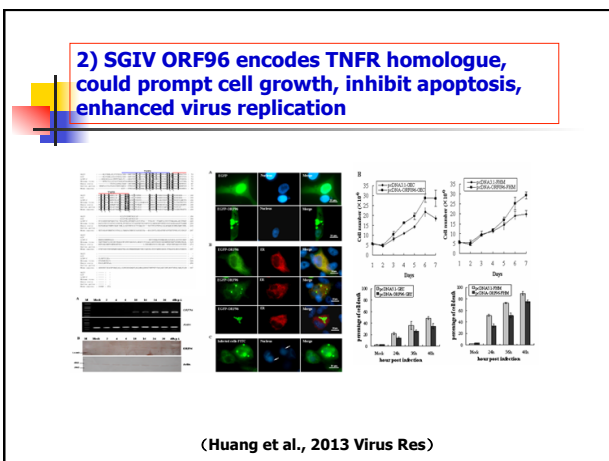
SGIV ICP46磷酸化分析

Xia et al., Arch Virol, 2009;
Xia et al., Virus Res, 2010.

3. SGIV could encode the genes homologous to host, and manipulate host immune response (Immune evasion)







3) SGIV ORF062R encodes IGF homologue. SGIV IGF is an E gene, localized in cytoplasm

16

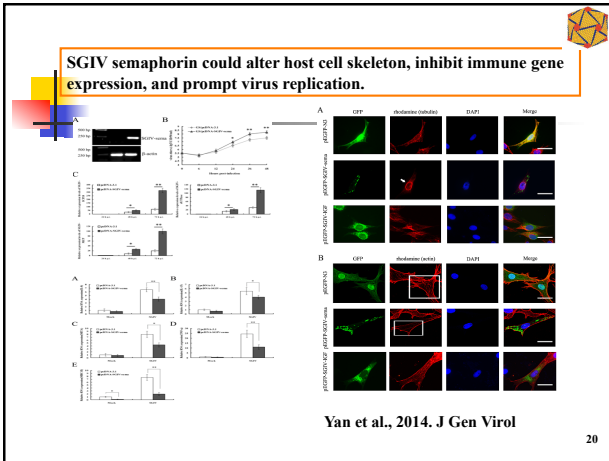
SGIV IGF could prompt cell cycle, improve host cell growth, enhance virus replication and virus induced apoptosis.

Yan et al., *J Gen Virol*. 2013 12:2759-70.

17

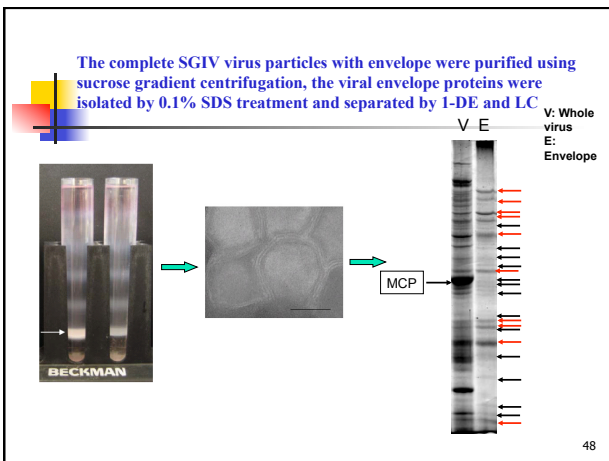
4) SGIV ORF155R encodes a semaphorin homologue. SGIV semaphorin is an E gene, localized in cytoplasm.

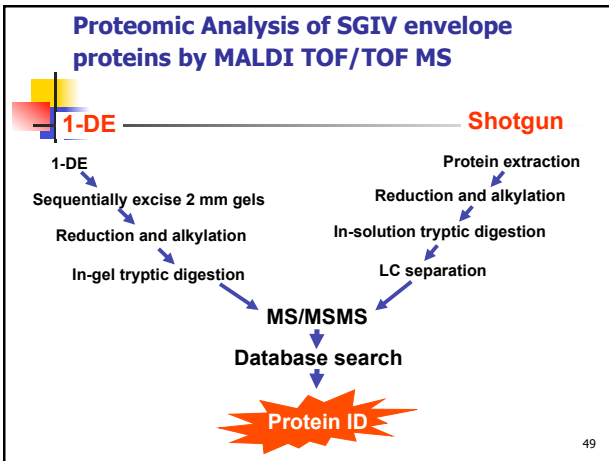
19

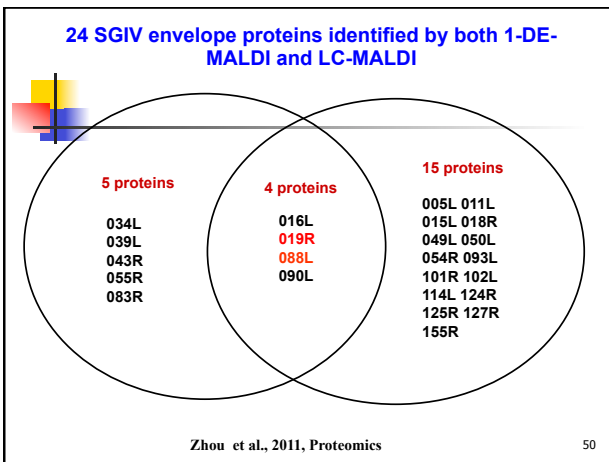


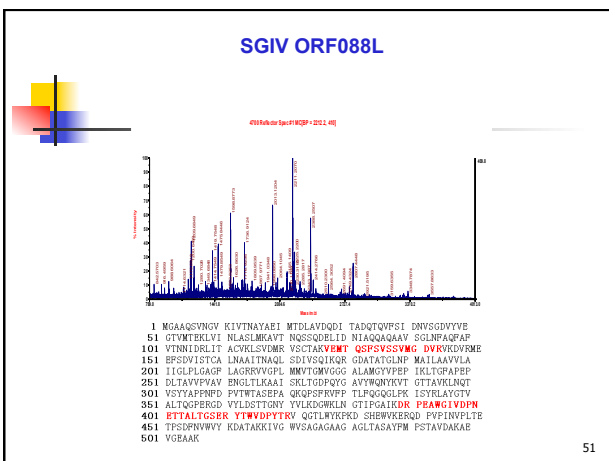
(VII) Functional proteomics of SGIV envelope proteins

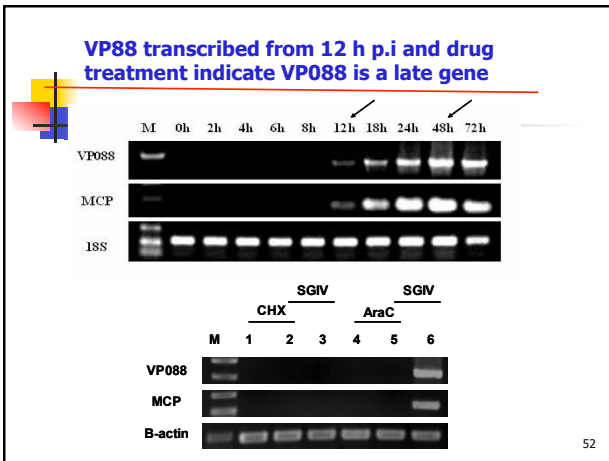
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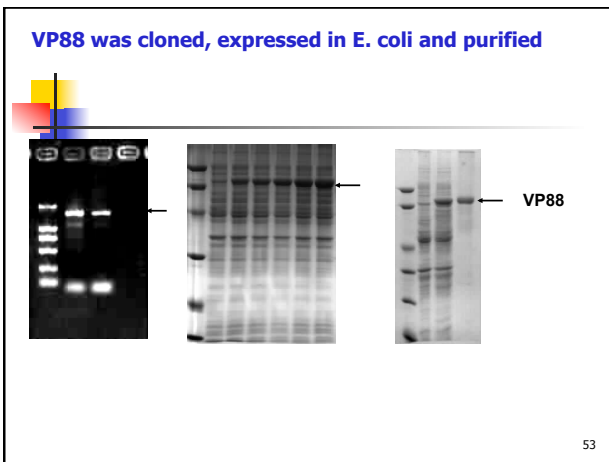


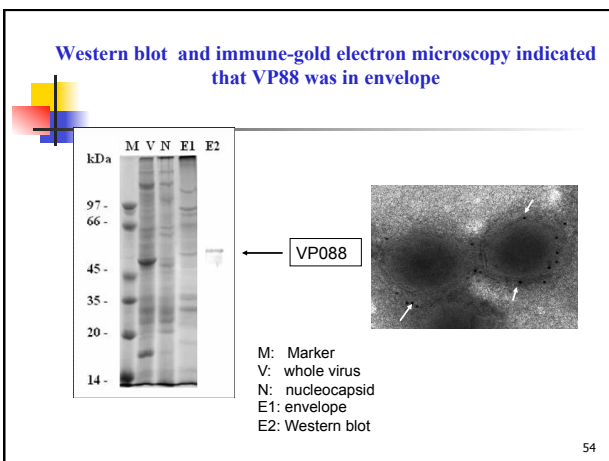




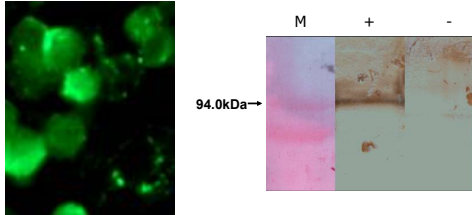








VP88 attached to the host cell membrane and reacted with a protein of 94 kDa

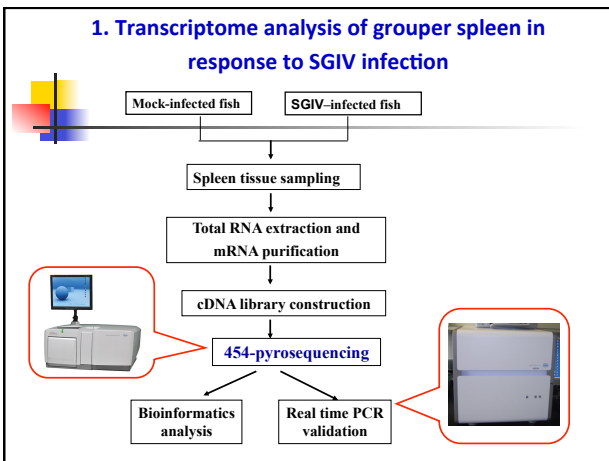


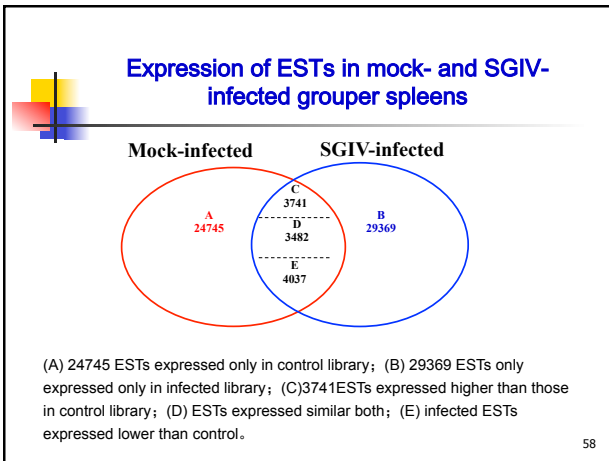
The figure consists of two parts. On the left is a fluorescence microscopy image showing green fluorescent spots on a dark background, representing VP88 attached to host cell membranes. On the right is a Western blot with three lanes labeled 'M', '+', and '-'. An arrow points to a band at 94.0 kDa in the '+' lane, indicating the presence of a specific protein. The 'M' lane is a molecular weight marker, and the '-' lane is a negative control.

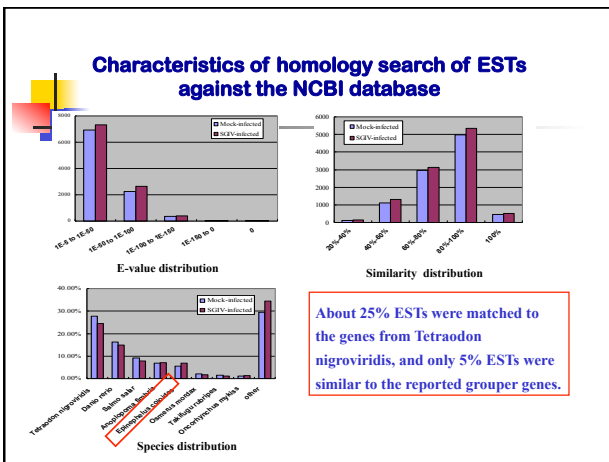
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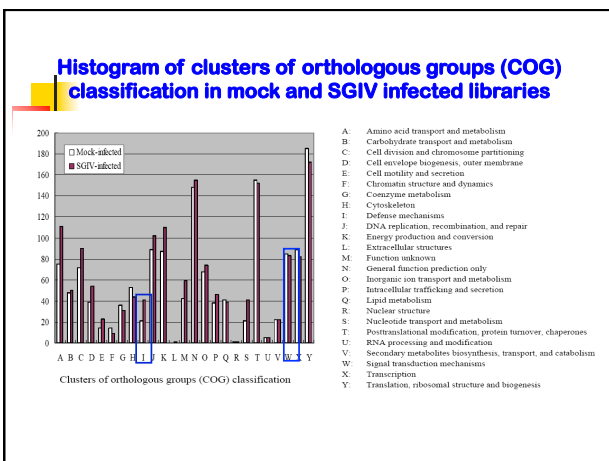
(VIII) Grouper, *Epinephelus coioides* immune responses to SGIV infection

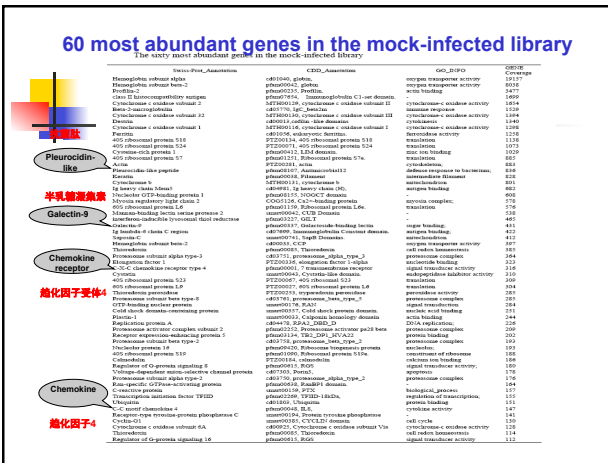
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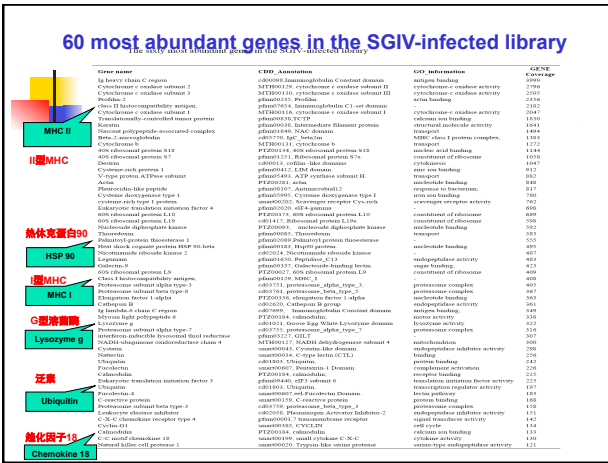




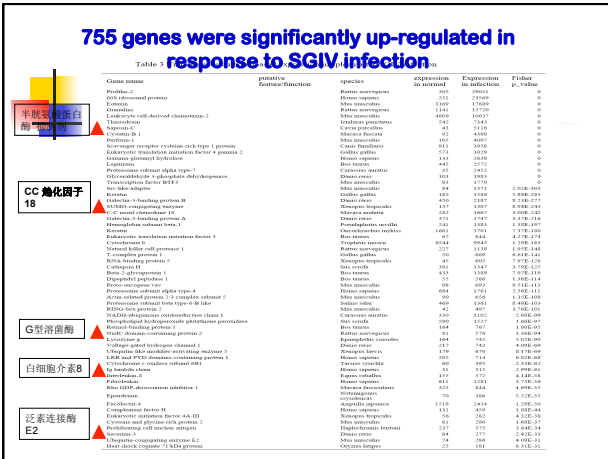




Gene	Expression
hsp70	1000
hsp90	800
hsp60	600
hsp40	400
hsp27	300
hsp25	250
hsp22	200
hsp18	150
hsp15	100
hsp12	80



Gene	Expression
hsp70	1000
hsp90	800
hsp60	600
hsp40	400
hsp27	300
hsp25	250
hsp22	200
hsp18	150
hsp15	100
hsp12	80

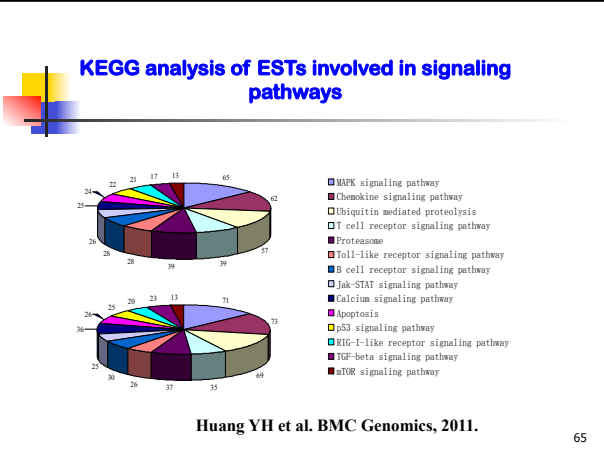


Gene	Expression
hsp70	1260
hsp90	1700
hsp60	1000
hsp40	600
hsp27	300

695 genes were significantly down-regulated in response to SGIV infection

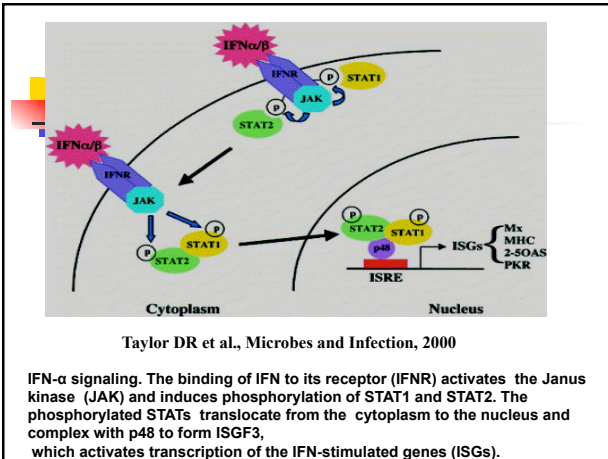
Table 4 Unique genes with decreased expression in spleen after SGIV infection

Gene ID	Gene Name	Protein Function	Species	Expression in spleen	Expression in spleen	Fold
				log2	log2	change
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2. Molecular cloning, expression and functional analysis of ISG15 from the grouper transcriptome

Huang XH et al. Fish & Shellfish Immunology, 2013.



Cloning and Bio-informatic analysis of EcISG15

Coding sequence (E. coli) (GenBank accession: F60515.1):

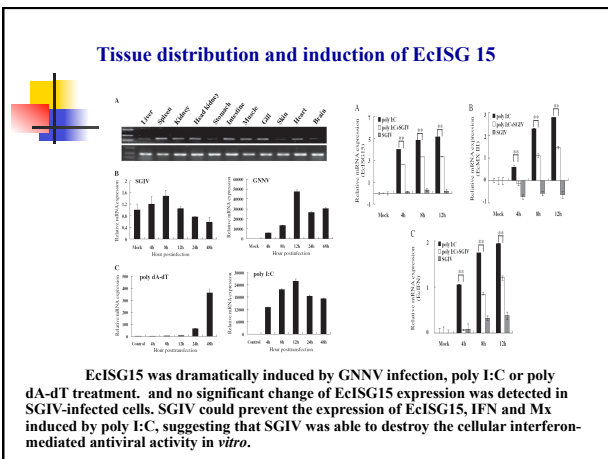
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MDSGKGGKAEAGYATGCGATGACACTTLE
GGKSTVAFKAPVDYPTITDAMKAGGAGAGCGTGTGKATATGATGATGATGATGGAGAGATGGAGGATGAT
...

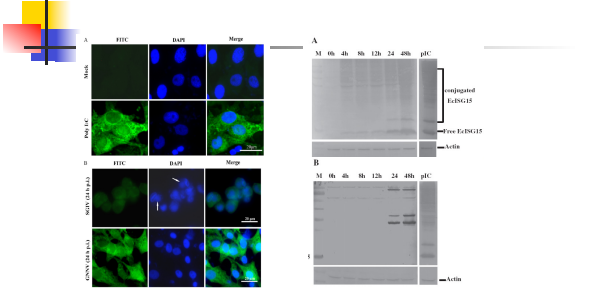
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Phylogenetic tree and domain analysis showing EcISG15 similarity to other ISG15 proteins from various species like *Sebastes schlegelii*, *Ospidirosetta diabolus*, etc.

EcISG15 encoded a polypeptide of 155 amino acids and shared the highest amino acid similarity to ISG15 of *Sebastes schlegelii*, and showed different identity to other known fish ISG15 homologs. EcISG15 shared the typical characteristics of known ISG15s with two UBL domains and a c-terminal LRGG motif.

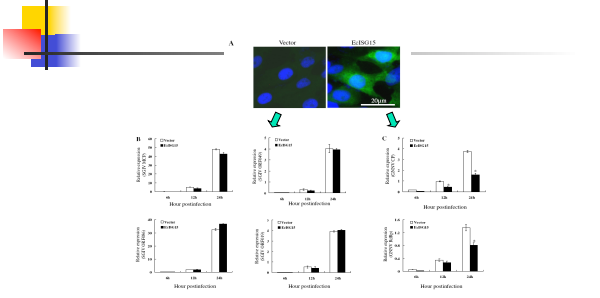


Subcellular localization and formation of EcISG15 conjugates



EcISG15 expression and ISGylation were evoked by GNVV infection or poly I:C stimulation, but not SGIV. The possibility of the absence of conjugation in SGIV infected-cells might be due to the disruption of IFN regulated response by SGIV infection *in vitro*.

The effects of EcISG 15 overexpression on SGIV and GNVV gene transcription



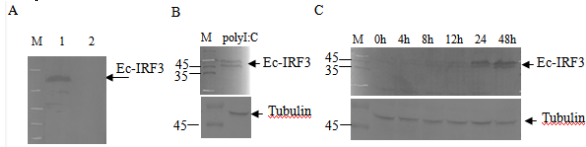
Overexpression of EcISG15 *in vitro* inhibited GNVV replication but not SGIV, suggesting that EcISG15 played the antiviral role via protein ISGylation in response to RNA virus infection.

3. Identification and functional characterization of grouper IRF3



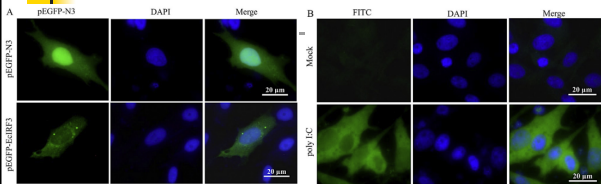
Huang et al., 2015, Fish Shellfish Immunol

Expression of rEc-IRF3 and antibody preparation



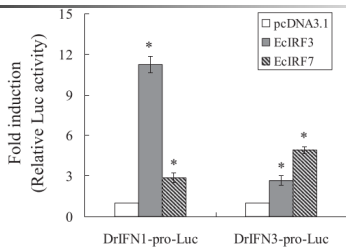
Western blotting assay showed that anti-Ec-IRF3 specially recognized not only the recombinant fused protein His-Ec-IRF3, but also the newly synthesized Ec-IRF3 proteins in poly I:C treated or RGNNV infected cells

Subcellular localization of Ec-IRF3 in grouper cells

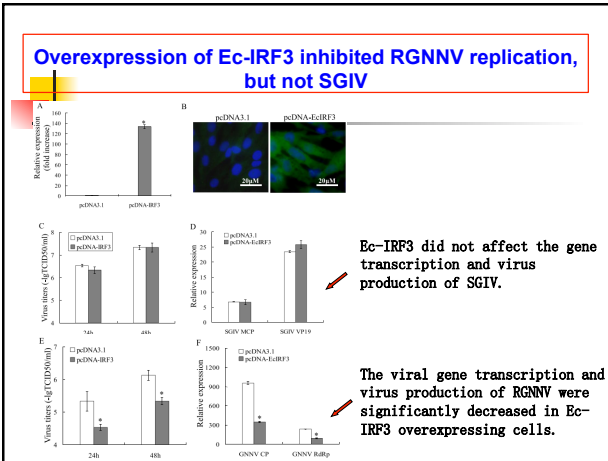


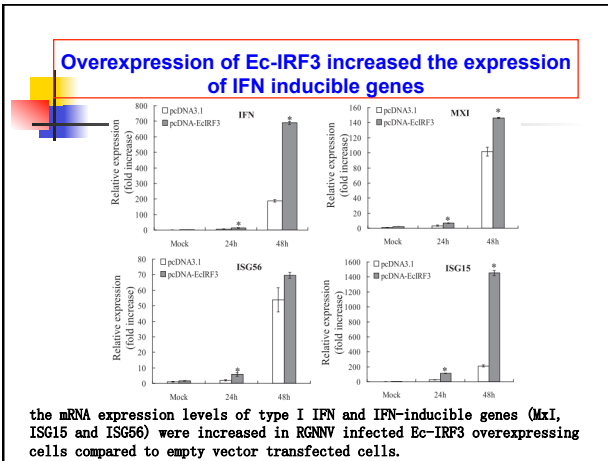
In pEGFP-Ec-IRF3 transfected cells, the green fluorescence was mostly observed in the cytoplasm. Immunofluorescence assay indicated that the fluorescence signal of Ec-IRF3 was increased significantly after infection with RGNNV or treatment with poly I:C, but moderately at the late stage of SGIV infection.

Induction of IFN promoter activity by Ec-IRF3



Reporter gene assay showed that EcIRF3 activated zebrafish type I IFN and type III IFN promoter *in vitro*, our results suggested that EcIRF3 might be involved in the regulation of IFN expression.

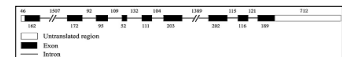




4. Identification and functional characterization of grouper IRF7

Cui et al., 2011, Dev Comp Immunol

Molecular cloning of *Epinephelus coioides* IRF7 gene

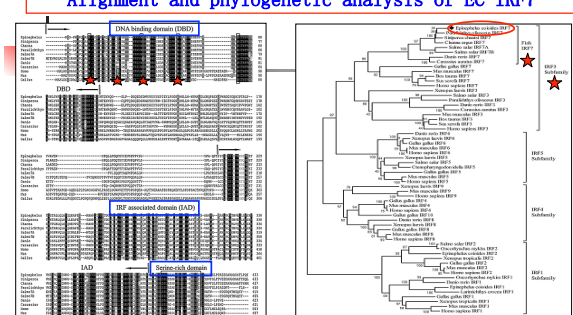


EC-IRF7 cDNA is of 2089 bp
5' UTR: 46 bp
Open reading frame (ORF): 1302 bp—433 aa
3' UTR: 741 bp
Four well-spaced tryptophan residues
Three mRNA instability motifs (ATTTA)

EC-IRF7 genomic DNA is about 5.7 kb
9 exons/8 introns & GT/AG rule



Alignment and phylogenetic analysis of EC-IRF7

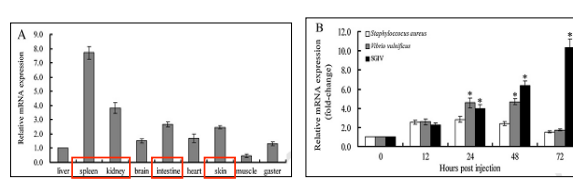


Conserved domains across species:
DBD, IAD, Serine-rich domain, Tryptophan residues

Vertebrate IRF7 subgroup, IRF3 subfamily.
Closer phylogenetic distance to other fish IRF7s



EC-IRF7 expression profiles in healthy and pathogens-stimulated fish



EC-IRF7 showed strong expression in immune-related tissues (spleen, kidney, intestine et al.);

EC-IRF7 was greatly up-regulated in spleen after stimulation with *Vibrio vulnificus* and Singapore grouper iridovirus (SGIV)



Expression of rEc-IRF7 and antibody preparation

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Ec-IRF7 activates fish type I IFN promoter

A IFN luciferase

Condition	Relative stimulation
Empty vector	~1.0
IRF7 100 ng	~1.5
IRF7 400 ng	~2.2
IRF7 600 ng	~2.8

B IFN luciferase

Condition	pDNA-ΔIRF7	pDNA-IRF7
Mock	~1.0	~2.5
SGIV	~1.0	~16.0

Ec-IRF7 was able to activate zebrafish IFN promoter in a dose-dependent manner; SGIV stimulation could enhance the activation activity of Ec-IRF7 on IFN promoter

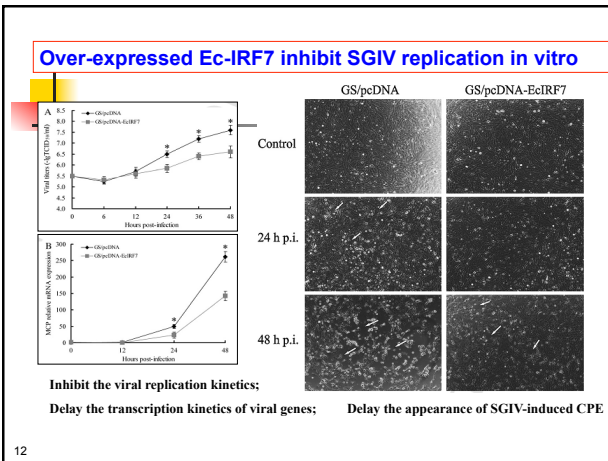
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Subcellular localization of Ec-IRF7

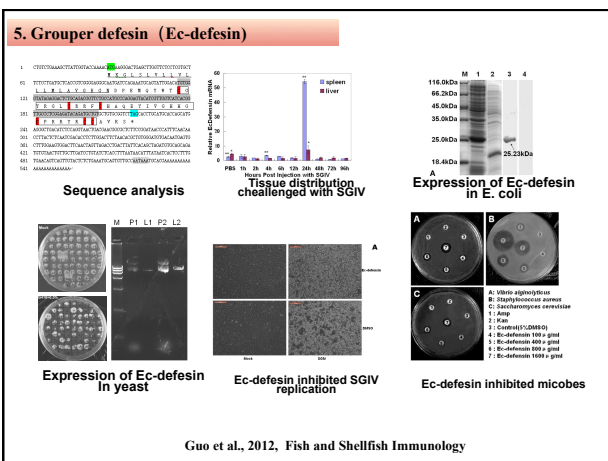
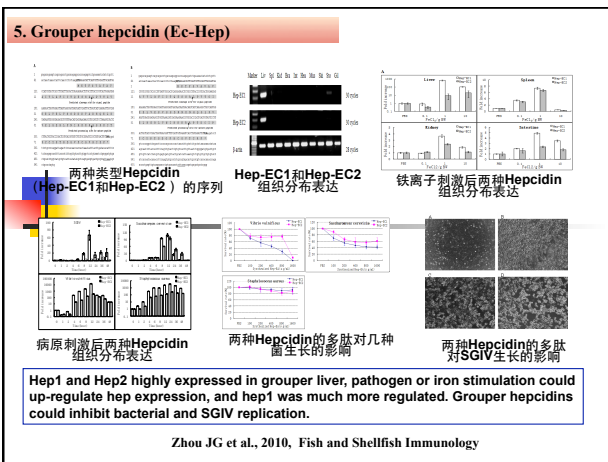
	FITC	DAPI	Merge
Negative serum			
Mock			
SGIV			

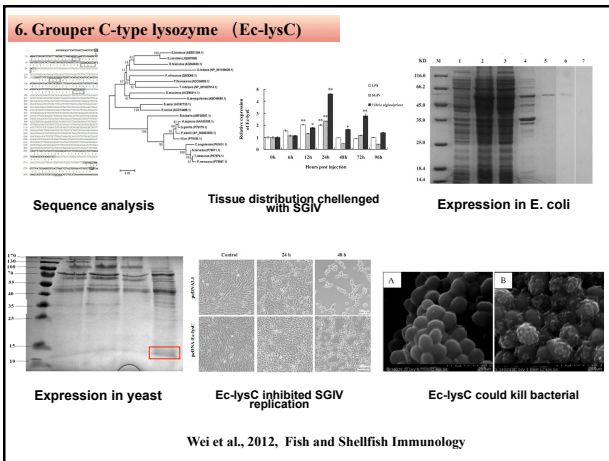
In non-stimulated cells, Ec-IRF7 resides in the cytoplasm; After SGIV infection, Ec-IRF7 translocated into the nucleus; (indicating Ec-IRF7 might be phosphorylated and activated during SGIV infection)

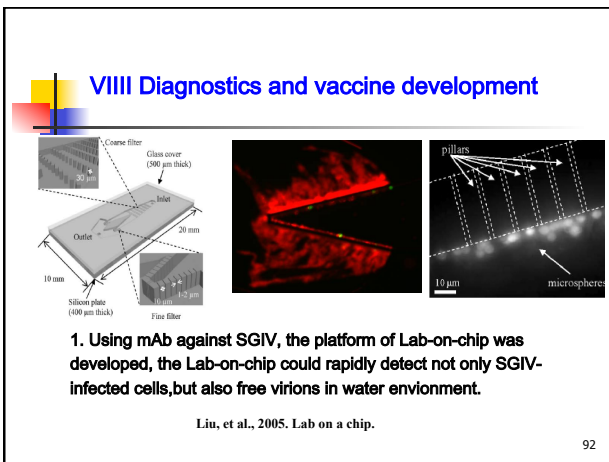
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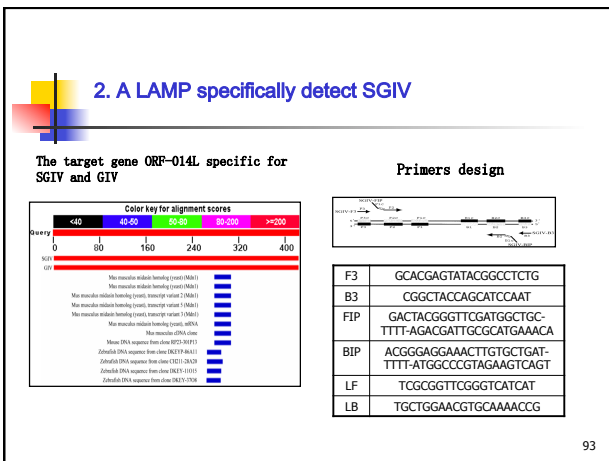


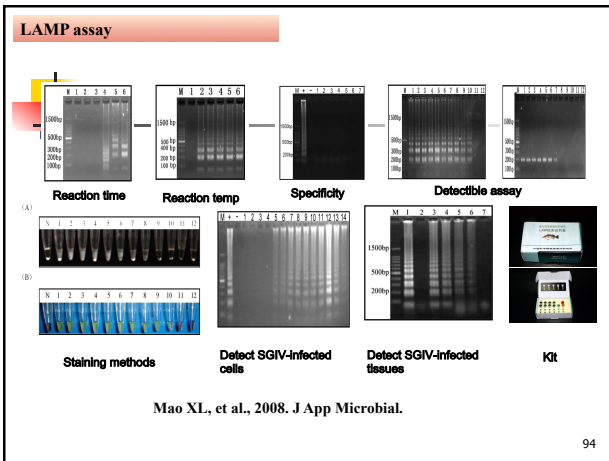
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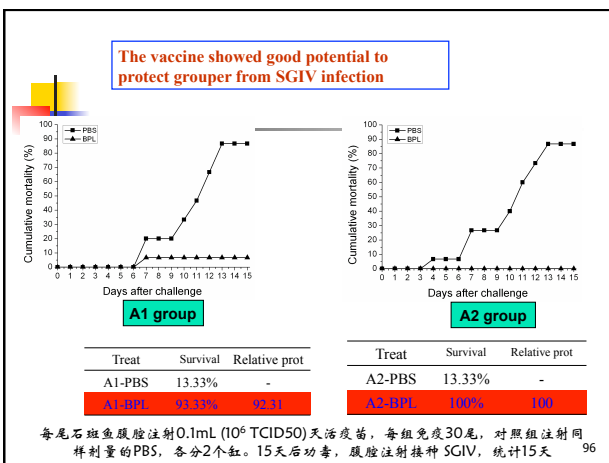
3. Inactivated vaccine against SGIV

Inactivated whole virus; BPL treated virus for 24h at 4° C

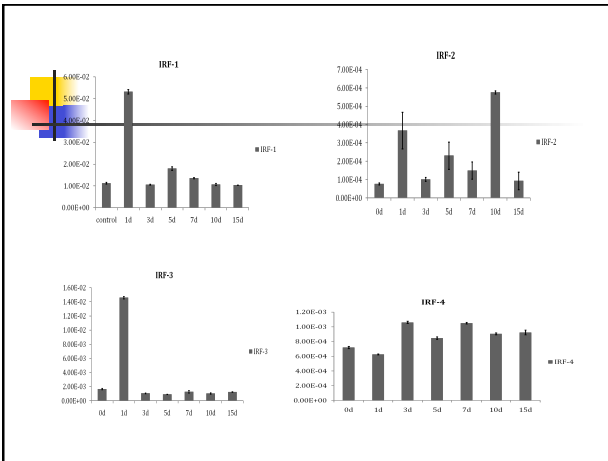
BPL 浓度	处理 时间	盲传三代结果		
		1	2	3
1: 250	6h	+	+	+
	12h	-	-	-
	24h	-	-	-
1: 500	6h	+	+	+
	12h	-	-	-
	24h	-	-	-
1: 1000	48h	+	+	+
	60h	-	+	+
	72h	-	+	+
	48h	+	+	+
1:2000	60h	+	+	+
	60h	+	+	+
	72h	+	+	+

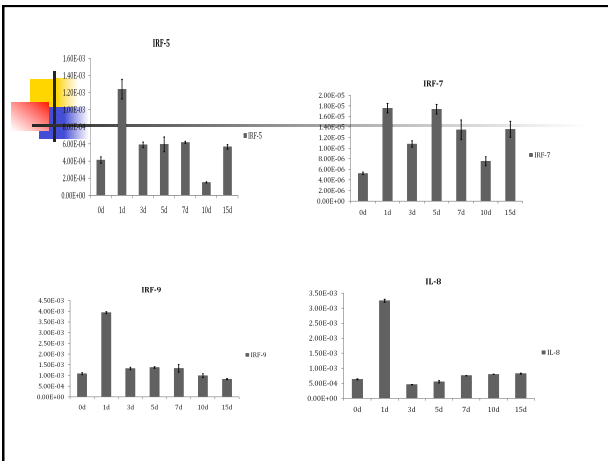
(+ positive CPE, -negative CPE)

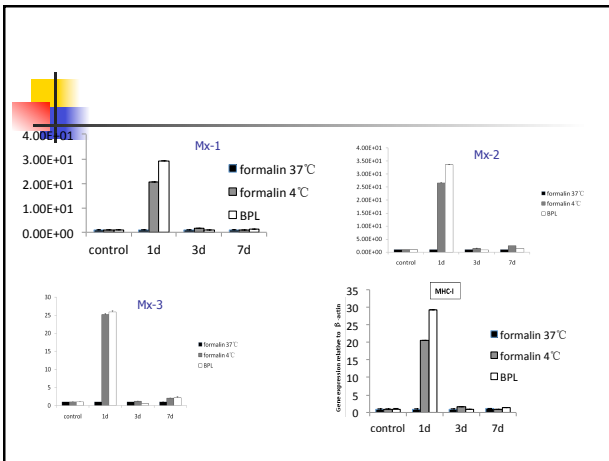
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Perspective Work

- ❖ Interaction & regulations between virus and host
- ❖ Development of vaccines and antiviral strategies
- ❖ Novel diagnostics

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Reference book

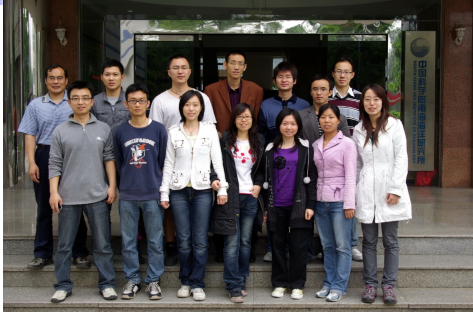
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