

On the road to disease: testing the stress-induced susceptibility hypothesis in amphibian populations adjacent to roads

Hall, E.M., Brunner, J.L., Hutzenbiler, B., Crespi, E.J.
School of Biological Sciences, Washington State University, Pullman WA



Photo: Alex Sherpak



Road Map

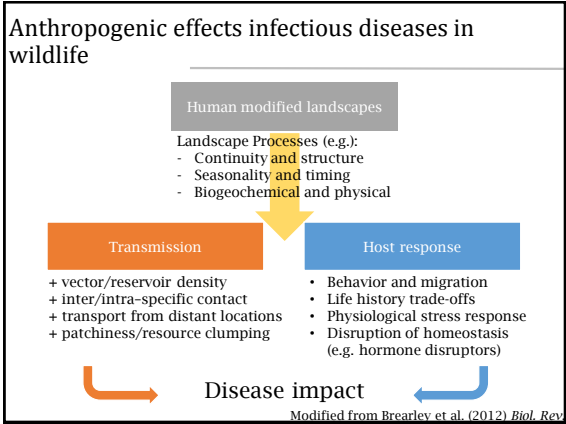
1. Why stress can increase susceptibility to disease
2. Combining surveillance (eDNA) and dose-response experiments
3. Understanding why variance within a species across an ecological context is important

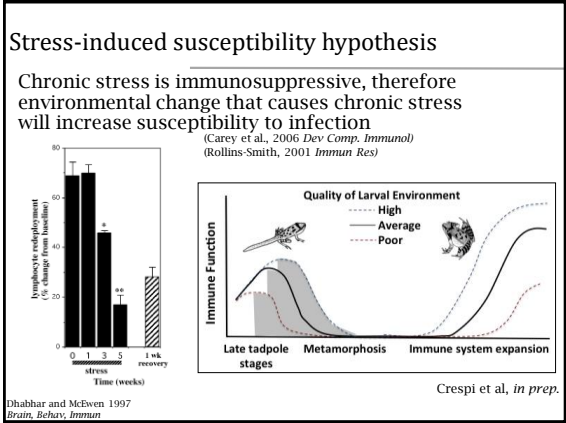
Anthropogenic effects infectious diseases in wildlife

53% (10/19) studies showing an increase in wildlife disease prevalence related to human-modified landscapes

Brearley et al. (2012) *Biol. Rev.*







Examples: stress-induced susceptibility in amphibians

Stressors that suppress immune function:

- **Glucocorticoids** (naturally during metamorphosis)
(e.g., Rollins-Smith 2001, *Immunol Res*; Belden and Kiesecker 2005, *J. Parasitol*, but see Searle et al 2014, *J Exp Zool*)
- **Toxicity** (Contaminants, agrochemicals, etc)
(e.g., Rohr et al 2008, *Nature*; Forson and Storfer 2006, *Ecol Appl*)
- **Nutritional deficit**
(Gervasi and Foufopoulos 2008, *Funct Ecol*; Venesky 2012, *Oecologia*)

A Belden and Kiesecker 2005

Group	Mean number of mesozoa (RE)
Control	~15
Ethanol	~15
Cort	~25

B

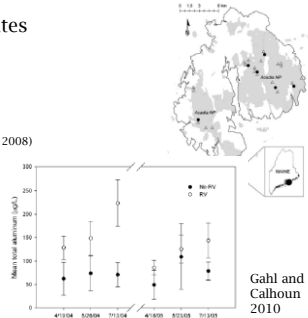
Group	Mean number of bacteria (RE)
Control	~35
Ethanol	~35
Cort	~10

Scale up to population level?

Examples: stress-induced susceptibility in amphibians

Environmental correlates with disease:

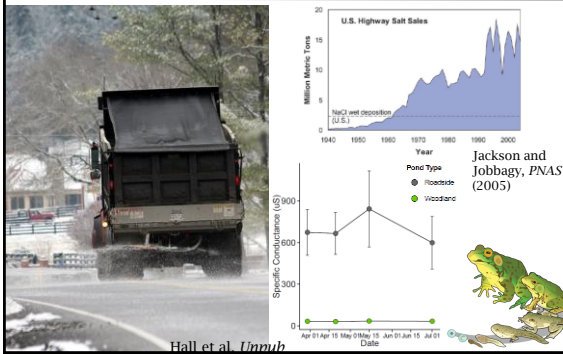
- Agricultural fields (e.g., Miller et al 2009)
- Urbanization/Industry (e.g., Skelly et al 2006, St. Amour et al 2008)
- Catchment position (Gahl and Calhoun 2010)
- Roads (Urban 2006, *Cons Bio*; Pauza et al 2010, *DOA*)



Gahl and Calhoun 2010

What is the mechanism?

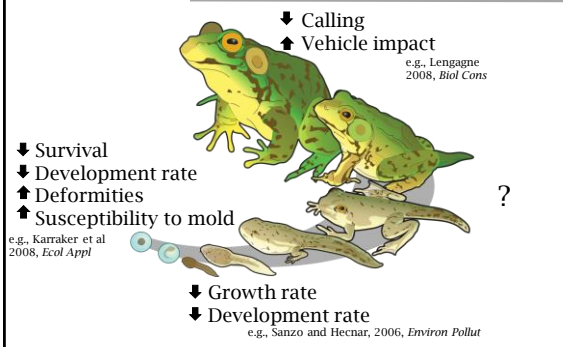
Roads are a major stressor to amphibians



Hall et al. *Ummh*

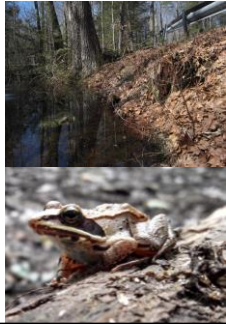
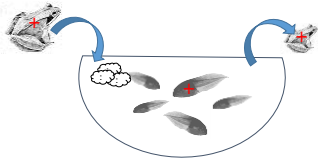
Jackson and Jobbagy, *PNAS* (2005)

Roads are a major stressor to amphibians

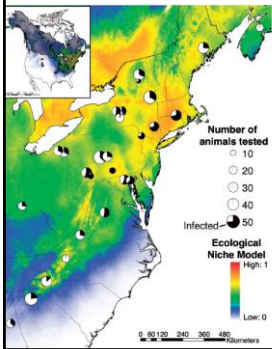


Wood frog system

- Explosive breeders
- Ephemeral fishless ponds
- Mortality from ranavirus reaches >90% of tadpoles, symptoms are observable



Wood frog system: ranavirus infection across the range



Found a higher prevalence of low-level infection in wood frog adults at center of range

- No correlation with stress hormones (GCs)
- Center of range is also highest density of human population

Crespi *et al* (2015), *ICB*

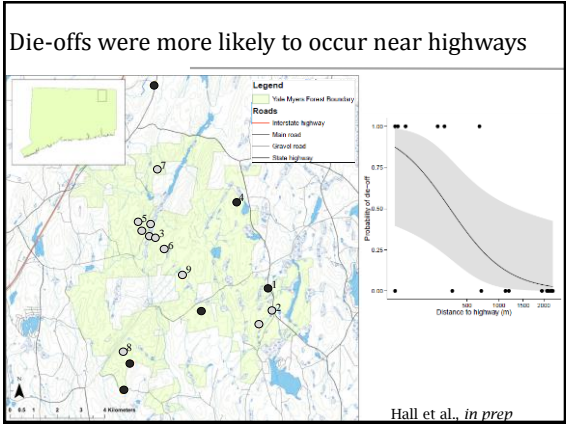
Testing the stress-induced susceptibility hypothesis

1. Are roads associated with ranavirus die-offs?

Host density, behavior, and physiology are affected by roadside conditions, thus ranavirus dynamics will vary by proximity to roads.



Approach:
Die-off and eDNA surveillance across a forested area dissected by roads



Wildlife disease surveillance: eDNA

eDNA is trace DNA in environmental samples. Mixture of degraded DNA from many different organisms. (Bohmann et al 2014, *Trends Ecol Evol*)

Strengths

- Detect multiple species with one sample (rare & cryptic)
- Relate pathogen and host densities
- Non-invasive, fast and easy
- High sensitivity

Weaknesses

- Not able to distinguish host species infected with pathogen
- High sensitivity prone to false positives
- Cannot distinguish between infectious/degraded pathogens

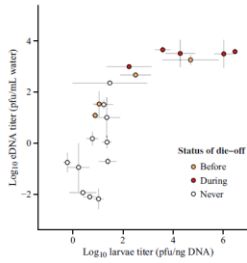
Using eDNA to detect ranaviruses in pond water

- Strengths: Not prone to false positives
- Weaknesses: Not as sensitive
- Some false negatives found -> sample more!

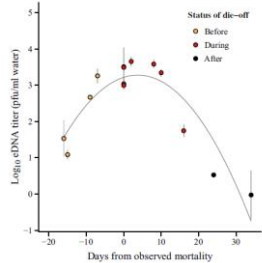
Sampled 20 wood frog ponds twice (before and after metamorphic climax) for eDNA (3 filters) and larvae (5 in 12 ponds)

Using eDNA to detect ranaviruses in pond water

RV eDNA titers reflected tadpole titers



RV titers peaked around die-off



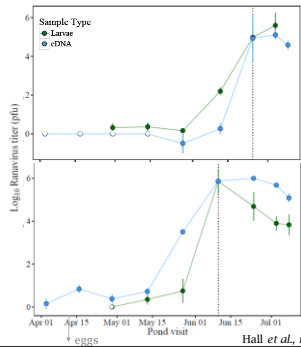
Hall et al., 2015, *Molecular Ecology Resources*

Using eDNA to detect ranaviruses in pond water

2014: Sampled 8 ponds every two weeks:

- 3 eDNA samples
- 10 larval samples
- Stages, densities, water chemistry

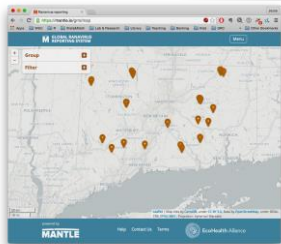
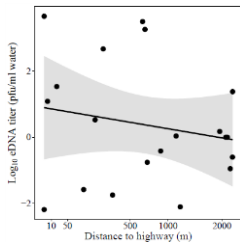
Monitored for die-offs every week (> 5 carcasses)



Hall et al., in prep

What's different about where die-offs occur?

Ranaviruses are ubiquitous...



O'Connor, K.M., T.A.G. Rittenhouse, and J.L. Brunner, in prep.

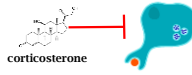
...so why are die-offs more likely in some ponds but not others?

Hall et al., in prep

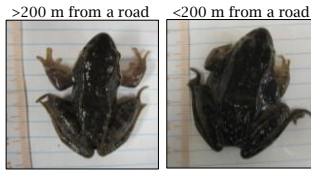
Specific hypotheses

How might road run-off increase susceptibility to infection?

1. Glucocorticoids may directly down regulate immune function, increasing susceptibility.



Adults (Hall et al., *in prep*)
 - Roadside had more bloating, and bloated adults had hormone profiles indicative of chronic stress

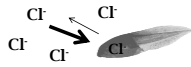


Tadpoles (Hall et al., *in prep*)
 - Higher baseline GCs in those collected from roadside ponds (when raised in freshwater or saltwater)

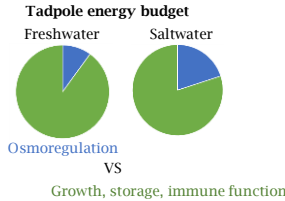
Specific hypotheses

How might road run-off increase susceptibility to infection?

2. Osmoregulation is costly and reduces the amount of energy available for fighting infection.



Tadpoles (Hall et al., *in prep*)
 - Slower growth in roadside ponds with higher salinity
 - Reduced feeding behavior when raised in road salt
 - Gill edema in road salt exposure in lab



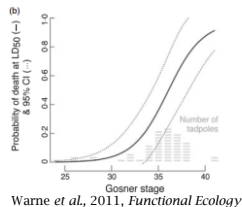
Specific hypotheses

How might road run-off increase susceptibility to infection?

3. Stress of poor conditions alters the timing of developmental traits associated with susceptibility.



Tadpoles (Hall et al., *in prep*)
 - Slower development rate in road salt exposure in lab
 - Greater variation in development rate in roadside ponds

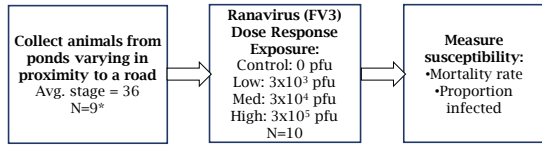


Warne et al., 2011, *Functional Ecology*

Testing the stress-induced susceptibility hypothesis

2. Do roads increase susceptibility to ranavirus infection?
Roadside conditions influence host physiology, thus individual susceptibility to infection will be related to proximity to roads.

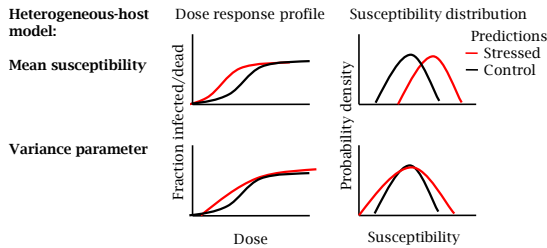
Approach: Dose-response exposure to FV3 ranavirus



*Collected from ponds that did not have RV die-offs at that time

The power of dose-response experiments

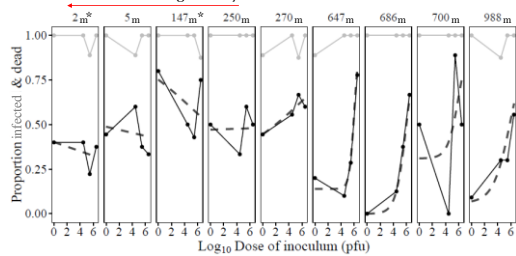
We can determine both the LD₅₀ and the distribution of susceptibility using a dose-response experiment



Modified from Ben-Ami 2010 *Am Nat*

Variation across and within populations

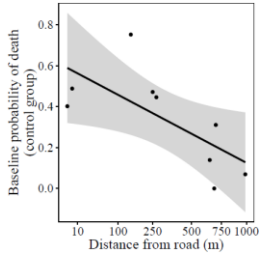
Despite high prevalence of infection across ponds, the response to a (secondary) challenge in the laboratory varied by proximity to a paved road (increasing salinity)



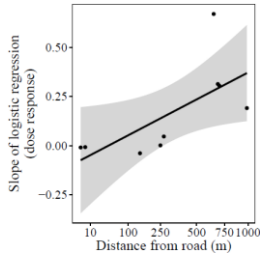
Hall et al., *in prep*

Variation across and within populations

Probability tadpoles will succumb to infection

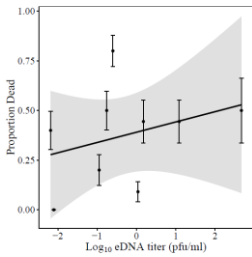


Variance in susceptibility to secondary infection

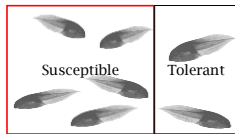


Hall et al., *in prep*

Potential explanations for differences in susceptibility

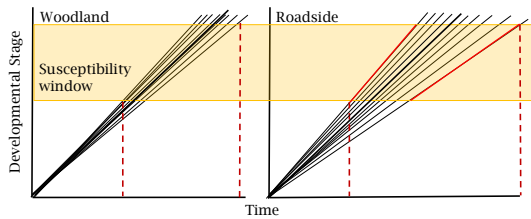


- ▲ Initial exposure
- ▲ Mortality to salinity
- Susceptible population already succumbed to infection




Potential explanations for differences in susceptibility

Roadside experimental enclosures had greater variance in development rate (Hall et al., *in prep*), and developmental stages vary in mortality to infection




Take home messages

1. Support for stress-induced (variance in) susceptibility (not as simple as it seems)
2. eDNA is a great tool for studying epidemiology
 - Can be related to titer in host community
 - Look at shedding rate and transmission
 - Non-invasive, minimal resources, easy to do
3. Dose-response experiments capture important factors
 - Can determine differences in lethal dose/mean susceptibility
 - Found interesting variance in susceptibility




Take home messages

1. Towards a mechanistic understanding of disease-associated declines
2. Combined lab and field experiments to strengthen associations between stressors and disease
3. Within-species differences in susceptibility across an ecological context is as important as among species



Findings and future directions

1. Does road salt stress alter transmission efficiency, immune function, or stress response to infection?
2. Does variance in development rate within a population increase the probability of a die-off?
3. Are there factors associated with the temporal pattern of when die-offs occur in a pond community?



Many Thanks

Committee: **Carl H. Elling Endowment**
Natural Resources Yale Myers Forest
 Dr. Erica Crespi **Conservation Endowment** Max Lambert
 Dr. Jesse Brunner **EPA STAR** Meredith Atwood
 Dr. Andrew Storfer Mark Ashton
 Dr. Jeb Owen

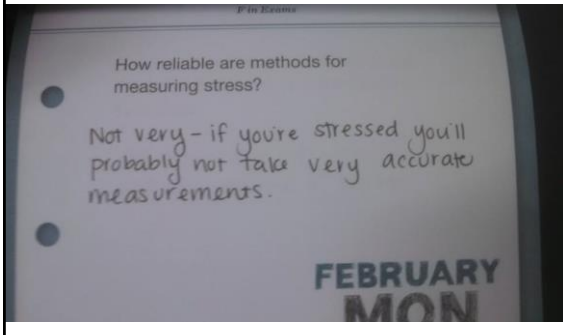
Lab manager:
 Jenn Cundiff

Collaborators:
 Dr. Dave Skelly
 Dr. Steve Brady
 Dr. Caren
 Goldberg



Undergraduates:
 Brandon
 Hutzenbiler
 Molly Diamond
 (CAS Undergraduate
 Summer Research
 Mini-grants)

Questions?



Literature Cited

Belden, L. K. and J. M. Kiesecker (2005). "Glucocorticosteroid hormone treatment of larval treefrogs increases infection by *Alaria* sp trematode cercariae." *Journal of Parasitology* 91(3): 686-688.

Bohmann, K., A. Evans, et al. (2014). "Environmental DNA for wildlife biology and biodiversity monitoring." *Trends Ecol Evol* 29(6): 358-367.

Brearley, G., J. Rhodes, et al. (2013). "Wildlife disease prevalence in human-modified landscapes." *Biol Rev Camb Philos Soc* 88(2): 427-442.

Crespi, E. J., L. J. Rissler, et al. (2015). "Geophisology of Wood Frogs: Landscape Patterns of Prevalence of Disease and Circulating Hormone Concentrations across the Eastern Range." *Integrative and comparative Biology* icv096.

Dhabhar, F. S. and B. S. Mowen (1997). "Acute stress enhances while chronic stress suppresses cell-mediated immunity in vivo: A potential role for leukocyte trafficking." *Brain, behavior, and immunity* 11(4): 286-306.

Forson, D. D. and A. Storfer (2006). "Atrazine increases ranavirus susceptibility in the tiger salamander, *Ambystoma tigrinum*." *Ecological Applications* 16(6): 2325-2332.

Gahl, M. K. and A. J. K. Calhoun (2010). "The role of multiple stressors in ranavirus-caused amphibian mortalities in Acadia National Park wetlands." *Canadian Journal of Zoology-Revue Canadienne De Zoologie* 88(1): 108-121.

Gervasi, S. S. and J. Foufopoulos (2008). "Costs of plasticity: responses to desiccation decrease post-metamorphic immune function in a pond-breeding amphibian." *Functional Ecology* 22(1): 100-108.

Hall, E. M., E. J. Crespi, et al. (2015). "Evaluating environmental DNA-based quantification of ranavirus infections in wood frog populations." *Molecular ecology resources*.

Jackson, R. B. and E. C. Jobling (2005). "From icy roads to salty streams." *Proceedings of the National Academy of Sciences of the United States of America* 102(41): 14487-14488.

Karraker, N. E., J. P. Gibbs, et al. (2008). "Impacts of road deicing salt on the demography of vernal pool-breeding amphibians." *Ecological Applications* 18(3): 724-734.

Langagne, T. (2008). "Traffic noise affects communication behaviour in a breeding anuran, *Hyla arborea*." *Biological Conservation* 141(8): 2023-2031.

Literature Cited

- Miller, D. L., M. J. Gray, et al. (2009). "Pathologic findings in larval and juvenile anurans inhabiting farm ponds in Tennessee, USA." *Journal of wildlife diseases* 45(2): 314-324.
- Pauza, M. D., M. M. Driessen, et al. (2010). "Distribution and risk factors for spread of amphibian chytrid fungus *Batrachochytrium dendrobatidis* in the Tasmanian Wilderness World Heritage Area, Australia." *Diseases of Aquatic Organisms* 92(2-3): 153-199.
- Kohr, J. R., A. M. Schotthoefer, et al. (2008). "Agrochemicals increase trematode infections in a declining amphibian species." *Nature* 455(7217): 1235-1239.
- Rollins-Smith, L. A. (2001). "Neuroendocrine-immune system interactions in amphibians - Implications for understanding global amphibian declines." *Immunologic Research* 23(2-3): 273-280.
- Sanzo, D. and S. J. Hecnar (2006). "Effects of road de-icing salt (NaCl) on larval wood frogs (*Rana sylvatica*)." *Environmental Pollution* 149(2): 247-256.
- Searle, C. L., L. K. Belden, et al. (2014). "Stress and chytridiomycosis: exogenous exposure to corticosterone does not alter amphibian susceptibility to a fungal pathogen." *Journal of Experimental Zoology, Part A: Ecological, Genetics, and Physiology* 321(5): 243-253.
- Skelly, D. K., S. R. Bolden, et al. (2006). "Urbanization and disease in amphibians." *Disease ecology, community structure and pathogen dynamics*: 153-167.
- St-Amour, V., W. M. Wong, et al. (2008). "Anthropogenic influence on prevalence of 2 amphibian pathogens." *Emerging infectious diseases* 14(7): 1175.
- Urban, M. C. (2006). "Road facilitation of trematode infections in snails of northern Alaska." *Conservation Biology* 20(4): 1143-1149.
- Venesky, M. D., T. E. Wilcoxen, et al. (2012). "Dietary protein restriction impairs growth, immunity, and disease resistance in southern leopard frog tadpoles." *Oecologia* 169(1): 23-31.
